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Production, characterization, and antigen specificity of recombinant 62-71-3, a candidate monoclonal antibody for rabies prophylaxis in humans

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ABSTRACT Rabies kills many people throughout the developing world every year. The murine monoclonal antibody (mAb) 62-71-3 was recently identified for its potential application in rabies postexposure prophylaxis (PEP). The purpose here was to establish a plant-based production system for a chimeric mouse-human version of mAb 62-71-3, to characterize the recombinant antibody and investigate at a molecular level its interaction with rabies virus glycoprotein. Chimeric 62-71-3 was successfully expressed in Nicotiana benthamiana. Glycosylation was analyzed by mass spectrometry; functionality was confirmed by antigen ELISA, as well as rabies and pseudotype virus neutralization. Epitope characterization was performed using pseudotype virus expressing mutated rabies glycoproteins. Purified mAb demonstrated potent viral neutralization at 500 IU/mg. A critical role for antigenic site I of the glycoprotein, as well as for two specific amino acid residues (K226 and G229) within site I, was identified with regard to mAb 62-71-3 neutralization. Pseudotype viruses expressing glycoprotein from lyssaviruses known not to be neutralized by this antibody were the controls. The results provide the molecular rationale for developing 62-71-3 mAb for rabies PEP; they also establish the basis for developing an inexpensive plant-based antibody product to benefit low-income families in developing countries.—Both, L., van Dolleweerd, C., Wright, E., Banyard, A. C., Bulmer-Thomas, B., Selden, D., Altmann, F., Fooks, A. R., Ma, J. K.-C. Production, characterization, and antigen specificity of recombinant 62-71-3, a candidate monoclonal antibody for rabies prophylaxis in humans. FASEB J. 27, 2055–2065 (2013). www.fasebj.org

Key Words: plant biotechnology · molecular pharming · PEP · tobacco

Rabies has the highest human case:fatality ratio of all infectious diseases, and it is widely accepted that there is no effective treatment after onset of symptoms (1–3). The causative agent is a negative-stranded RNA virus in the order Mononegavirales, family Rhabdoviridae, genus Lyssavirus (4). All mammals are susceptible and can transmit rabies virus (RV; ref. 5), and both canine and sylvatic (wildlife) circulation patterns are recognized (6). Most human exposures are associated with the bites of rabid animals, in particular, unvaccinated dogs, and transmission of RV in their saliva (7). Human rabies cases have also been attributed to probable aerosol exposures in laboratories or airborne exposures in caves with high densities of bats (8, 9). In addition, atypical transmission through butchering and processing of rabid animals, as well as human-to-human transmission by organ or tissue transplantation have been reported (10, 11).

Although viral spread to the central nervous system (CNS) and result in encephalitis are almost invariably

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Abbreviations: ELISA, enzyme-linked immunosorbent assay; ERIG, equine rabies immunoglobulin; FAVN, fluorescent antibody virus neutralization; HRIG, human rabies immunoglobulin; HRP, horseradish peroxidase; LBV, Lagos bat virus; mAb, monoclonal antibody; OD, optical density; PEP, post-exposure prophylaxis; PNA, pseudotype neutralization assay; RIG, rabbits immunoglobulin; RP-ESI-MS, reverse-phase electrospray ionization mass spectrometry; RV, rabies virus; scFv, single-chain variable fragment;

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fatal, the disease is preventable through postexposure prophylaxis (PEP). Swift administration of PEP is virtually 100% effective in preventing the onset of symptoms and fatal clinical disease after exposure (12–17). Rabies PEP is based on 3 pillars: wound cleansing, administration of rabies vaccine, and infiltration of rabies immunoglobulins (RIGs) of either human or equine origin (HRIGs or ERIGs, respectively). However, insufficient access to RIGs restricts the administration of appropriate PEP across the developing world where the vast majority of the annual 55,000–70,000 rabies fatalities occur (18–22). To overcome the short supply and the safety issues with blood-derived RIG products, several human and murine monoclonal antibodies (mAbs) are being investigated (23–25). A recent report by the World Health Organization (WHO) Rabies Collaborating Centres described the identification of three novel combinations of mAbs to replace RIGs (6). Stringent criteria concerning the neutralizing activity, binding specificities to different epitopes, immunoglobulin isotype, and history of hybridomas were used to evaluate the suitability of several murine mAbs. Combinations of 2 mAbs, all including mAb 62-71-3, were assessed both in vitro and in vivo and were shown to have an equal or superior efficiency to HRIGs in the hamster PEP model (6).

The objective of the present study was to clone and express a chimeric (mouse-human) full-length IgG1 version of mAb 62-71-3, using plants as an inexpensive production alternative to existing mammalian systems, and to perform a detailed molecular characterization of the recombinant mAb. Initially, a phage-displayed single-chain variable fragment (scFv) of mAb 62-71-3 was expressed in Escherichia coli and tested to confirm that the sequences for heavy and light chains correctly encoded for an antibody with neutralizing potency toward the virus. A chimeric 62-71-3 full-length IgG was then cloned, expressed, and purified from Nicotiana benthamiana leaves. The plant-derived mAb was investigated using mass spectrometry for glycan analysis, RV glycoprotein enzyme-linked immunosorbent assay (ELISA), fluorescent antibody virus neutralization (FAVN) and pseudotype neutralization assay (PNA). Mutations in antigenic site I of the virus were also performed to confirm cloning of the correct variable regions of mAb 62-71-3 were finally generated by immunizing BALB/c mice with the vaccine strain ERA (6, 31). The cDNA sequences for the constant regions of a human mAb IgG1 κ, using the primers listed in Table 1 and the cloning strategy described in Supplemental Data. A schematic representation of the cloning strategy is shown in Fig. 1A.

Cloning and expression of the 62-71-3 phage-displayed scFv

mAb 62-71-3 is a hybridoma-derived IgG2b antibody, originally generated by immunizing BALB/c mice with the rabies vaccine strain ERA (6, 31). The cDNA sequences for the variable regions of mAb 62-71-3 were received from Apotech (Lausanne, Switzerland). To confirm cloning of the correct variable region sequences, an scFv version of mAb 62-71-3 was initially expressed in E. coli. The variable regions of heavy and light chains were amplified by PCR and were connected with a flexible 15aa linker, using the primers listed in Table 2 and the cloning strategy described in Supplemental Data. A schematic representation of the cloning strategy is shown in Fig. 1B. The heavy and light chain sequences of the chimeric 62-71-3 were then subcloned from the entry vector pDONR into the Gateway destination vector pEAQ-HT-Dest3 for expression in plants (32) or with the Gateway destination vectors pcDNA-Dest40 and pEF-Dest51 (Invitrogen, Carlsbad, CA, USA) for expression in mammalian cells. Agrobacterium tumefaciens cultures (strain LBA4404) transformed with either the heavy-chain or light-chain vectors were each adjusted to optical density at 600 nm (OD600 nm) = 1 by diluting the cells with infiltration buffer (10 mM MES and 10 mM MgCl₂, pH 5.6) and combined. The cells were incubated in the dark (2 h, room temperature) before infiltration of N. benthamiana plants with a 1-ml syringe without needle (2–3 leaves/plant). Soluble leaf extracts were prepared by grinding leaf tissue in a mortar and centrifugation.

The HEK293-derived mAb 62-71-3 was generated by mixing 1 μg of the heavy- and light-chain plasmids with Fugene 6 (Roche, Basel, Switzerland) according to manufacturer’s instructions and transfection of 70% confluent 293T-17 cells in a 6-well plate containing 2 ml medium/well (DMEM plus 15% FBS and 1% Pen/Strep). The plates were incubated at 37°C (5% CO₂), and medium was changed after 24 h. After

### MATERIALS AND METHODS

#### Cloning and expression of chimeric mAb 62-71-3

The murine variable regions of mAb 62-71-3 were grafted onto the constant regions of a human mAb (IgG1 κ), using the primers listed in Table 2 and the cloning strategy described in Supplemental Data. A schematic representation of the cloning strategy is shown in Fig. 1B. The heavy and light chain sequences of the chimeric 62-71-3 were then shuffled from the entry vector pDONR into the Gateway destination vector pEAQ-HT-Dest3 for expression in plants (32) or with the Gateway destination vectors pcDNA-Dest40 and pEF-Dest51 (Invitrogen, Carlsbad, CA, USA) for expression in mammalian cells. Agrobacterium tumefaciens cultures (strain LBA4404) transformed with either the heavy-chain or light-chain vectors were each adjusted to optical density at 600 nm (OD600 nm) = 1 by diluting the cells with infiltration buffer (10 mM MES and 10 mM MgCl₂, pH 5.6) and combined. The cells were incubated in the dark (2 h, room temperature) before infiltration of N. benthamiana plants with a 1-ml syringe without needle (2–3 leaves/plant). Soluble leaf extracts were prepared by grinding leaf tissue in a mortar and centrifugation.

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2–3 d, the supernatants were collected, passed through a 0.45-μm filter, and stored at 4°C.

**SDS-PAGE and Western blot analysis**

The soluble fraction of a plant extract was passed through Miracloth (EMD Millipore, Billerica, MA, USA) and a 0.45-μm filter, and the mAb was purified by protein G (Sigma, Gillingham, UK) affinity chromatography. Both the purified mAb and the crude plant extract were analyzed by SDS-PAGE and semidy Western blot analysis. SDS-PAGE was performed using the In Vitrogen Migiel system and In Vitrogen NuPage buffers. Electrophoresis was carried out in 4–12% gradient gels (In Vitrogen), which were stained with Coomassie brilliant blue or subjected to Western blotting. Transfer to the membrane (Amersham Hybond-ECL; Amersham Biosciences, Little Chalfont, UK) was carried out in a semidy system (Invitrogen). The membrane was blocked with milk (3%), incubated with horseradish peroxidase (HRP)-coupled antibodies at a concentration of 1:10,000, and developed (Amersham ECL Plus Western blotting detection kit, Amersham Hyperfilm ECL).

**RV glycoprotein ELISA**

A commercial ELISA kit (Bio-Rad Platelia Kit; Bio-Rad, Hemel Hempstead, UK) was used to investigate recombinant antibody binding to its target antigen. The ELISA is based on RV glycoprotein coated on the plate and detection of antibody using HRP-coupled protein A. Clarified plant extract supernatants (at 3, 5, and 7 d postinfiltration) were applied to the wells, and the assay was run according to the manufacturer’s instructions.

The RV glycoprotein ELISA was also used for competition experiments, as described recently (33), with minor modifications. Briefly, plates were incubated with saturating amounts of purified plant-derived mAbs 62-71-3 and E559 for 1 h at room temperature. For the initial production of the plant-derived mAb E559, the E559 hybridoma was obtained from Dr. Thomas Müller [Friedrich Loeffler Institut (FLI), Wusterhausen, Germany; ref. 6]. The hybridoma heavy- and light-chain sequences for this murine IgG1 mAb were biotinylated with the No-Weigh Sulfo-NHS-LC-Biotin (Pierce, Rockford, IL, USA), according to manufacturer’s instructions, and 50 μl of biotinylated mAb E559 (2.5 μg/ml) was then added to each well, incubated for 5 min at room temperature, and rinsed 5 times with 100 μl of Tris-buffered saline with 0.1% Tween (TBST). Subsequently, wells were incubated for 1 h at room temperature with 50 μl of a 1:5,000 dilution of streptavidin-HRP (Sigma). Wells were rinsed, and HRP activity was detected by the addition of 3,3’,5,5’-tetramethylbenzidine dihydrochloride substrate (Sigma). Color development was allowed to proceed for 10 min at room temperature before the addition of 2 M H2SO4 to terminate the reaction. The OD was measured at 450 nm on an ELISA plate reader.

**Generation of lentiviral pseudotype viruses**

DNA plasmids encoding the HIV gag-pol, luciferase reporter gene, and lyssavirus glycoproteins were used as previously reported (33). Briefly, pseudotype viruses were generated by mixing 1 μg of the HIV gag-pol plasmid, 1 μg of the glycoprotein plasmid, and 1.5 μg of the reporter gene plasmid with Fugene 6 (Roche) and transfection of 293T-17 cells in a 6-well plate containing 2 ml medium/well (DMEM plus 15% FBS and 1% Pen/Strep). The plates were incubated at 37°C (5% CO2), and medium was changed after 24 h. After 2 d, the supernatants containing pseudotype virus were collected, passed through a 0.45-μm filter, and frozen at −80°C.

**PNA**

The concentration of the purified plant-derived mAb was measured with the bicinchoninic acid (BCA) protein assay kit (Pierce). The phage-displayed 62-71-3 scFv or plant-derived 62-71-3 IgG (5 μg/ml) was added to medium (DMEM plus 10% FCS and 1% Pen/Strep) and titrated in doubling dilutions across a 96-well plate, starting with a 1:10 or 1:20 dilution (final volume of 50 μl/well). Controls were cells only, virus only, and cells and virus, which were set up with appropriate amounts of medium. Pseudotype virus (50 μl) was added to each well (apart from cells-only control) at a dilution where the international reference serum (OIE serum) neutralizes 100% at 1:40 serum dilution (~100 median tissue culture infective dose (TCID50)). The plate was centrifuged (500 rpm, 5 s) and incubated at 37°C (5% CO2) for 1 h. Medium (100 μl) containing 2 × 10⁴ BHK cells was added to each well (apart from virus-only control), making up a total volume of 200 μl/well. The plate was centrifuged (500 rpm, 5 s) and incubated at 37°C (5% CO2) for 48 h. Medium (115 μl) was removed from each well, and 75 μl BrightGlO (Promega, Madison, WI, USA) was added. After measuring absolute infection (light units) by the pseudotype virus with a
luminometer, the relative neutralization was calculated, and titration curves were generated.

**FAVN assay**

The standard FAVN test was set up in a similar way to the PNA, starting with a 1:8 dilution of antibody. OIE\(^+\) and OIE\(^-\) sera were included as controls for CVS. To assess neutralization of other lyssaviruses, a modified FAVN was performed as described previously (63). Briefly, for each sample, mAbs and the viruses were incubated at 37°C (5% CO\(_2\)) for 1 h before adding 4 × 10\(^5\) BHK cells to each well. The plates were then incubated at 37°C (5% CO\(_2\)) for 48 h, fixed in 80% acetone, and air-dried. The staining was carried out by adding 50 μl of fluorescein isothiocyanate (FITC)-conjugated antibody (Cen
tor, Philadelphia, PA, USA), specific for the RV nucleoprotein, to each well. After a 30-min incubation at 37°C (5% CO\(_2\)), each plate was washed 3 times with PBS. Excess PBS was removed by briefly inverting the microplates on absorbent paper, and the neutralizing titer was evaluated by fluorescent microscopy.

**Glycan analysis of the plant-derived mAb 62-71-3**

A glycoproteomic analysis was undertaken by in-gel digestion of 5-carbamidomethylated sample and analysis by reverse-phase electrospray ionization mass spectrometry (RP-ESI-MS), as described previously (63). Tandem MS results were also subjected to Mascot MS/MS ion search (Matrix Science, London, UK; http://www.matrixscience.com).

**Mutational analysis of the RV glycoprotein**

Chimeric RV/Lagos bat virus (LBV) glycoproteins were generated by swapping antigenic sites I–IV of the RV strain CVS11 (accession no. EU352767) with those of LBV strain Nig56–E559, known to be directed against antigenic site II of the viral glycoprotein.

**RESULTS**

**Characterization of phage-displayed 62-71-3 scFv**

The cDNA sequences for the variable regions of the heavy and light chains of mAb 62-71-3 were cloned into a phagemid vector (Fig. 1A) to verify that the sequences correctly encoded for an antibody with neutralizing potency toward the virus. The phage-displayed 62-71-3 scFv was tested for its neutralization of a lentivirus (HIV) pseudotyped with the RV glycoprotein (strain CVS11) and demonstrated potent, dose-dependent neutralization of the pseudotype virus (Fig. 2A), confirming that the correct heavy and light chains were cloned. Controls included a nonspecific scFv (Fig. 2A), as well as cells only, virus only, and cells and virus (data not shown).

**Production and antigen-binding properties of plant-derived chimeric mAb 62-71-3**

After the initial neutralization experiments with the 62-71-3 scFv, a chimeric antibody was constructed (Fig. 1B) and expressed in *N. benthamiana*. Infiltrated *N. benthamiana* leaves showed robust expression of 62-71-3 IgG (estimated as ~3–4% of total soluble protein by
The breadth of neutralization was further investigated using lentiviruses pseudotyped with a panel of lyssavirus glycoproteins from different phylogroup I viruses and some related Eurasian lyssaviruses. In addition to the CVS11 strain, the plant-derived IgG demonstrated potent neutralization of several different lyssaviruses, including ERA (RV), RV1787, and RV634 (Fig. 3A, B). Diminished neutralization was observed for the two Duvenhage virus strains SA06 and RV131 (Fig. 3B) and the two European bat lyssavirus type 1 strains RV9 and RV20 (Fig. 3A). The three Eurasian lyssaviruses Irikut, Khujand, and Aravan were all strongly neutralized (Fig. 3C). We also tested RV neutralization by the plant-derived purified mAb with the FAVN assay. The plant-derived antibody demonstrated a neutralizing titer of ~500 IU/mg for strain CVS11, while no neutralization was observed for LBV, an RV-related phylogroup II virus (negative control).

Glycoproteomic analysis

Sequence analysis of heavy and light chains of mAb 62-71-3 predicted the presence of a single potential N-linked glycosylation site in the antibody Fc region. The plant-derived antibody was subjected to glycoproteomic analysis by RP-ESI-MS. Glycopeptides comprising the Fc glycosylation site EEQYNSTYR (N-linked glycosylation site is underscored) were identified at the early retention time characteristic for this peptide (44, 63). The glycan analysis revealed that mAb 62-71-3 displayed glycan compositions typical of plant glycoproteins, with predominantly complex type glycans containing xylose and fucose, which are presumed to be the β1,2-linked xylose residues attached to the β-linked mannose and the α1,3-fucose residue linked to the Asn-linked N-acetyl-glucosamine (Fig. 4). Tandem MS results were subjected to Mascot MS/MS ion search, which confirmed the sample to contain essentially mAb 62-71-3.
Competition ELISA

To verify that the binding of mAb 62-71-3 does not depend on antigenic site II, the immunodominant epitope in mice, a competition experiment with the site II-specific mAb E559 was performed, using a protocol adopted from Marissen et al. (34). RV glycoprotein ELISA plate wells were incubated with saturating amounts of purified plantibodies before adding biotinylated hybridoma-derived mAb E559. As expected, the binding of biotinylated E559 was blocked when plates were preincubated with plant-derived mAb E559 (Fig. 5). In contrast, binding was not blocked when the 62-71-3 plant-derived antibody or no antibody (buffer only, negative control) were added, indicating that mAb 62-71-3 does not compete with the site II-specific mAb E559.

Mutational analysis of the RV glycoprotein

To investigate the antibody-antigen interaction in more detail, a set of RV glycoprotein mutants was analyzed regarding their neutralization by the 62-71-3 scFv. These mutants were based on CVS11 pseudotype viruses, each containing a separate replacement of one of the 4 major antigenic sites by the corresponding region from a phylogroup II virus (LBV.Nig56-RV1). The neutralization experiments with these mutated pseudotypes showed that neutralization by the 62-71-3 scFv was severely diminished when antigenic site I was altered (Fig. 6A). In contrast, mutations in other antigenic sites did not diminish neutralization by the 62-71-3 scFv. A similar experiment was undertaken with the plant-expressed 62-71-3 IgG (Fig. 6B). To demonstrate that the effects of mutating antigenic site I were specific for neutralization by mAb 62-71-3, we also included the plant-derived mAb E559, which targets antigenic site II of the viral glycoprotein (6). This chimeric version of mAb E559 was also cloned and expressed in planta (unpublished results) and purified from plant leaves by protein G affinity chromatography, for a direct comparison with the plant-derived purified 62-71-3 IgG. While mAbs E559 and 62-71-3 both showed potent and complete neutralization of a rabies pseudotype virus containing the CVS11 wild-type glycoprotein (CVS control), only mAb 62-71-3 showed diminished neutralization of the pseudotype virus containing the CVS11 glycoprotein mutated in site I, and a 100% neutralizing titer could not be defined for this mutant.

Amino acid residues present in antigenic site I were then compared for the lyssavirus pseudotypes assessed. Antigenic site I comprises 6 amino acid residues at positions 226–231 of the mature viral glycoprotein (without signal peptide). Alignments of the antigenic site I sequences from the different glycoproteins used in Fig. 3 and 4 revealed conservation of the 3 core amino acids L227, C228, and G229 (Table 3). However, whereas lyssavirus genotypes containing the residue K226 within antigenic site I were completely neutralized by mAb 62-71-3, lyssaviruses containing R226 showed diminished neutralization, and no 100% neutralizing titer could be defined for this point mutant (Fig. 6C). To confirm the critical role of residue K226, we investigated the 2 plant-derived mAbs 62-71-3 and...
E559 regarding their neutralization of pseudotypes containing glycoproteins with a K226R (lysine to arginine) mutation (Fig. 6D). Controls included pseudotypes containing the CVS11 wild-type glycoprotein or containing the CVS11 glycoprotein harboring a nonspecific mutation in antigenic site III residue N336, which has previously been found to be critical for neutralization by mAbs targeting site III (24, 57). This residue was mutated to the serine found in LBV (N336S) and had no effect on neutralization by mAb E559 (Fig. 6E). The expression levels of mAb 62-71-3 were estimated by quantitative ELISA and yielded ~3–4% of total soluble protein. This is ~100 mg/kg fresh tissue weight. Although these expression levels were satisfactory for the current study and do not necessarily require any additional improvements for large-scale production, further enhancement of expression would be desirable and might be achieved by codon-optimization and by using different expression vectors (two strategies that we are currently further investigating for mAb 62-71-3).

The original murine 62-71-3 IgG was converted here into a chimeric mAb with human antibody constant region sequences in order to improve half-life and reduce immunogenicity in humans. The resulting chimeric 62-71-3 IgG was shown to bind to its antigen in a functional ELISA and to neutralize a rabies pseudotype virus, confirming correct assembly and functionality of the antibody. We also confirmed the neutralizing activity of the plant-derived mAb in the FAVN assay and observed strong RV neutralization (500 IU/mg), similar to the previously reported potency of its murine hybridoma-derived counterpart (6).

To further corroborate the strong neutralizing potency by the plant-derived purified mAb 62-71-3, a panel of different pseudotype viruses was tested. Several lyssavirus pseudotypes were strongly neutralized, with

### DISCUSSION

Rabies occurs mainly among low-income families in Africa and Asia, and an inexpensive antibody product would be highly desirable for implementing appropriate PEP. Because of high costs and short supply, replacement of plasma-derived HRIGs and ERIGs remains a priority (35, 36), and a useful alternative is offered through the development of mAbs directed against the RV glycoprotein (37, 38). To overcome the short supply of RIGs, the WHO Rabies Collaborating Centres previously set out to investigate several mAbs for inclusion into an antibody cocktail (6, 35). In a WHO consultation in 2002, the mAbs 62-71-3, E559, 1112-1, D8, M777, and M727 were initially proposed (35). mAb 1112-1 was eventually excluded because of intellectual property reasons (6), and mAb D8 was excluded due to insufficient strain coverage (unpublished results). The remaining mAbs were shown to target the same epitope on the viral glycoprotein, namely antigenic site II, with the exception of mAb 62-71-3 (6). Three novel antibody cocktails were designed, all containing mAb 62-71-3 as an essential component (6).

In the present study, we report the cloning of a chimeric IgG version of mAb 62-71-3 and expression in a plant system. Initially, a phage-displayed scFv version of mAb 62-71-3 was produced and tested for its neutralizing potency against a rabies pseudotype virus. This step confirmed that the variable region sequences correctly encoded for an antibody with neutralizing potency toward the virus. A full-length chimeric antibody was then cloned and expressed in N. benthamiana. The expression levels of mAb 62-71-3 were estimated by quantitative ELISA and yielded ~3–4% of total soluble protein. This is ~100 mg/kg fresh tissue weight. Although these expression levels were satisfactory for the current study and do not necessarily require any additional improvements for large-scale production, further enhancement of expression would be desirable and might be achieved by codon-optimization and by using different expression vectors (two strategies that we are currently further investigating for mAb 62-71-3).

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To further corroborate the strong neutralizing potency by the plant-derived purified mAb 62-71-3, a panel of different pseudotype viruses was tested. Several lyssavirus pseudotypes were strongly neutralized, with

### Table 3. Alignment of antigenic site I residues of the tested lyssaviruses

<table>
<thead>
<tr>
<th>Strain</th>
<th>Neutralization</th>
<th>Site I</th>
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<tbody>
<tr>
<td>CVS (RV)</td>
<td>+</td>
<td>KLCGVL</td>
</tr>
<tr>
<td>ERA (RV)</td>
<td>+</td>
<td>KLCGVL</td>
</tr>
<tr>
<td>RV131 (DUVV)</td>
<td>–</td>
<td>RLCGIS</td>
</tr>
<tr>
<td>SA06 (DUVV)</td>
<td>–</td>
<td>RLCGIS</td>
</tr>
<tr>
<td>RV9 (EBLV1)</td>
<td>–</td>
<td>RLCGVP</td>
</tr>
<tr>
<td>RV20 (EBLV1)</td>
<td>–</td>
<td>RLCGVP</td>
</tr>
<tr>
<td>RV1787 (EBLV2)</td>
<td>–</td>
<td>RLCGIS</td>
</tr>
<tr>
<td>RV634 (ABLV)</td>
<td>+</td>
<td>RLCGIS</td>
</tr>
<tr>
<td>Irkat</td>
<td>+</td>
<td>KLCGMA</td>
</tr>
<tr>
<td>Aravan</td>
<td>+</td>
<td>KLCGVV</td>
</tr>
<tr>
<td>Khujand</td>
<td>+</td>
<td>KLCGVS</td>
</tr>
</tbody>
</table>

See Fig. 3. Asterisks below alignments indicate conserved residues. Neutralization: + indicates complete (100%) neutralization; – indicates that 100% neutralization could not be determined.
the exception of two Duvenhage strains (SA06 and RV131) and two European bat lyssavirus type 1 strains (RV9 and RV20). These results are also consistent with previous observations of mAb 62-71-3 produced in hybridoma cells (6), which suggested that the hybridoma-derived mAb broadly neutralizes different genotypes but is ineffective at neutralizing the genotypes Duvenhage virus and European bat lyssavirus type 1.

The purified recombinant mAb was then subjected to mass spectroscopy and was shown to be glycosylated with typical plant complex glycan structures. Although mammalian and plant cells share similar cellular machinery for antibody assembly, their post-translational modifications, including the processing of glycans, are not completely identical. While the processing in the ER is conserved among all eukaryotic species and is restricted to oligomannose type N-glycans, the processing in the Golgi varies between species, resulting in differences between plant-derived proteins and their mammalian counterparts (39). Plant glycans usually do not contain sialic acid residues and may contain typical plant complex glycan structures. Although mammalian N-glycans, the process-

interaction, a mutational analysis of the RV glycoprotein was undertaken. Lafon et al. (45) initially proposed an antigenic structure for the RV glycoprotein, defining several major epitopes. Antigenic site I has been defined as an epitope complex between residues 226 and 231 (46, 34), originally identified using a single mAb, which binds to an epitope, including residue 251 within antigenic site I (45, 46). Antigenic site II comprises a discontinuous epitope, including residues 34–42 and residues 198–200 (47). This antigenic site is targeted by most murine antibodies and mutations within these residues can result in heavily reduced pathogenicity of the virus (47). Antigenic site III comprises residues 330–338 and harbors two charged residues, K330 and R333, which are linked to viral pathogenicity and neuroinvasion (48, 49). Antigenic site IV contains overlapping linear epitopes with key residues 251 and 264 (50, 51). In addition, a few minor epitopes have been identified, e.g., between residues 342 and 343 (minor site a) and between residues 14 and 17 (45, 52).

Most studies on the antibody epitopes within the RV glycoprotein have been undertaken by sequencing viral escape mutants or by competition assays (53, 54). A few other studies have relied on investigating the interaction between antibodies and peptides derived from the viral glycoprotein, e.g., peptides derived by cleaving the glycoprotein with cyanogen bromide (55), short peptides expressed in yeast (56), as well as synthetic peptides for PEP-SCAN experiments (34). In the present study, we made use of pseudotype viruses containing chimeric RV/LBV glycoproteins generated by swapping the antigenic sites of RV strain CVS11 (phylogroup I lyssavirus) with those of LBV strain Nig56-RV1 (phyl-

**Figure 4.** The purification process was as follows: A) Western blot of purified recombinant mAb 62-71-3 scFv, B) Western blot of purified recombinant mAb 62-71-3 IgG, C) Mass spectroscopy of recombinant mAb 62-71-3 scFv, D) Mass spectroscopy of recombinant mAb 62-71-3 IgG.
group II virus). It is well established that neutralizing antibodies (including mAb 62-71-3) targeting phylogroup I viruses are not effective at neutralizing phylogroup II viruses (57, 58). The neutralization assays indicated that virus neutralization by the 62-71-3 scFv and the plant-derived 62-71-3 IgG specifically involve antigenic site I.

To examine more closely how differences in antigenic site I residues account for the differences in neutralization observed with different lyssavirus pseudotypes, their glycoprotein sequences were compared. The alignment revealed diverse antigenic site I residues of lyssaviruses belonging to phylogroup I and related Eurasian lyssaviruses (59–61). Although lyssaviruses containing the antigenic site I residue K226 (lysine) were completely neutralized by mAb 62-71-3, lyssaviruses containing the residue R226 (arginine) demonstrated diminished neutralization. To obtain further insight into the role of antigenic site I residues K226 or R226, we tested the 2 plant-derived mAbs 62-71-3 and E559 regarding their neutralization of pseudotypes containing glycoproteins with a K226R mutation. These experiments corroborated the critical role of K226 for neutralization by mAb 62-71-3. Moreover, pseudotypes containing a G229E mutation, which has previously been found in viral escape mutants resistant to a mAb targeting antigenic site I (34), showed diminished neutralization.

Previous studies indicated that mAb 62-71-3 was able to neutralize viral escape mutants of mAb E559 (directed against site II; ref. 6), similar to another mAb (mAb D1) known to target antigenic site III. This observation led the researchers (6) to suggest that mAb 62-71-3 targets an epitope different from antigenic site II, possibly antigenic site III, as it is well established that the vast majority of mAbs bind to sites II or III (48). Unfortunately, no viral escape mutants could be generated for mAb 62-71-3 in vitro (6), so that its epitope has been unknown so far. While this previous study (6) already suggested that the epitope of mAb 62-71-3 does not overlap with the epitope (site II) of mAb E559, our present study provides the first systematic epitope analysis. Relatively few antibodies specific for antigenic site I have been described to date (47, 59), and the neutralization assays undertaken in our current study demonstrate that antigenic site I, including residue K226 is critical for neutralization mAb 62-71-3. The demonstration of the binding specificity of mAb 62-71-3 to the RV glycoprotein at a molecular level provides an important rationale for including mAb 62-71-3 in RV-neutralizing antibody cocktails. The results provided here also indicate that mAb 62-71-3 would be a particularly good complement to antibodies targeting site II, such as mAb E559, and that mAbs specific to site III could now also be considered for inclusion in an antirabies cocktail product.

The neutralization experiments with pseudotypes containing mutated glycoproteins cannot differentiate between direct and indirect effects, i.e., it is currently not clear whether the diminished neutralization observed with pseudotypes harboring site I mutations is due to the absence of the targeted epitope or due to indirect/allosteric effects (i.e., certain amino acid residues in site I might mask neighboring epitopes that might be the real target of mAb 62-71-3). Nevertheless, our results explain why mAb 62-71-3 is relatively ineffective at neutralizing certain genotypes, in particular, those belonging to phylogroup II. Notably, many of these particular genotypes have very little clinical relevance, so the lack of neutralization of these genotypes does not pose a significant problem in the further development of the plant-derived mAb 62-71-3.

Future work should now focus on the evaluation of additional mAbs regarding their potential to complement mAb 62-71-3. The development of a combination of mAbs targeting distinct, nonoverlapping epitopes and covering a broad panel of RV isolates would be highly desirable. This study has focused on the molecular characterization of mAb 62-71-3, but two other features of this study targeted facilitating the clinical development of this antibody. First, we have focused on the partial humanization of mAb 62-71-3. Although using a murine mAb in PEP is not contraindicated, the use of human sequences is generally preferred. Second, we have explored a powerful plant expression system to generate the recombinant mAb. For almost 2 decades, the potential of plants for production of mAbs has been proposed (62). Recent advances in regulatory approval of plant biotechnologies and manufacturing scaleup now take this potential much closer to reality. There is now a real prospect of producing mAbs for rabies PEP in plants to exploit low costs, massive scalability, and potential technology transfer to resource-poor regions where rabies affects low-income families.

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