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## Synthesis and biological evaluation of ferrocene-based cannabinoid receptor 2

### ligands

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**Abstract.** Ferrocene analogues of known fatty acid amide hydrolase inhibitors and CB<sub>2</sub> ligands have been synthesized and characterized spectroscopically and crystallographically. The resulting bioorganometallic isoxazoles were assayed for their effects on CB<sub>1</sub> and CB<sub>2</sub> receptors as well as on FAAH. None had any FAAH activity but compound **3**, 5-(2-(pentyloxy)phenyl)-*N*-ferrocenylisoxazole-3-carboxamide, was found to be a potent CB<sub>2</sub> ligand ( $K_i = 32.5$  nM).

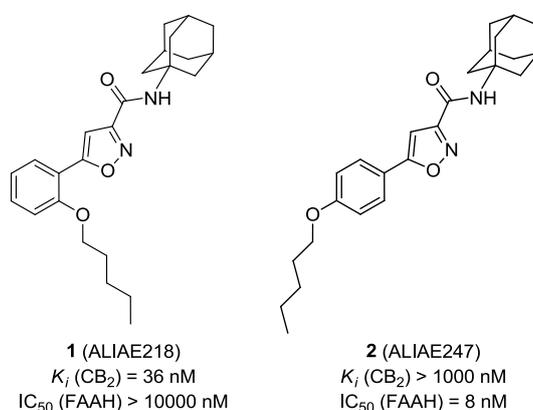
*Keywords:* isoxazole, ferrocene, cannabinoids, FAAH.

## ***Introduction***

*N*-Arachidonylethanolamine (AEA), also known as anandamide, is an endogenous signaling mediator of the endocannabinoid system.[1] It is formed from a membrane precursor, termed *N*-arachidonoyl phosphatidylethanolamine, through multiple biosynthetic pathways.[2, 3] It has been identified that there is a basal amount of AEA in tissues. The equilibrium and circulation of AEA are regulated by several relevant synthases, hydrolases, and transporters.[4] Fatty acid amide hydrolase (FAAH) was demonstrated to be responsible for the main hydrolysis of AEA into arachidonic acid and ethanolamine.[5] There is evidence that increasing AEA levels can produce anti-inflammation, pain relief, injury repair, and neuroprotection by activating cannabinoid receptors CB<sub>1</sub>/CB<sub>2</sub>. [6-8] Therefore, stimulating CB<sub>1</sub>/CB<sub>2</sub> receptors or inhibiting FAAH, thereby accumulating AEA, have been regarded as interesting therapeutic strategies for the treatment of pain, inflammation, nerve injury, or addiction.[4] Moreover, CB<sub>2</sub> inverse agonists have shown anti-inflammatory and anti-osteoporotic therapy in a manner dependent upon the migration of immune cells. [9-13]

Over recent decades, the development of CB<sub>2</sub> ligands and FAAH inhibitors has significantly progressed underpinned by numerous clinical trials of CB<sub>2</sub> agonists and FAAH inhibitors for the treatment of pain, inflammation, and central nervous system (CNS)-dependent disorders.[4, 14, 15] Recently, our work has identified 3-carboxamido-5-aryl-isozazole as a versatile scaffold for the design of CB<sub>2</sub> ligands

or FAAH inhibitors.[16-19] Of note, compound **1** (ALIAE218, Figure 1) showed potent affinity toward CB<sub>2</sub> receptors in the nanomolar range and significantly reduced dextran sulfate sodium (DSS)-induced colitis in mice.[16] Interestingly, the change of the pentoxy group of **1** from the ortho to the para position (**2**, ALIAE247, Figure 1) triggered a biological response switch from a CB<sub>2</sub> ligand to a FAAH inhibitor. Furthermore, the study of **2** in a model of DSS-induced colitis in mice identified its ability to alleviate inflammation.[18]



**Figure 1.** Structures of the known selective CB<sub>2</sub> ligand **1** and FAAH inhibitor **2**.

## Discussion

Ferrocene, known as an organometallic sandwich compound, has been used for drug design due to its suitable lipophilicity ( $\log P = 2.66$ ) facilitating membrane permeability, rotatable aromatic cyclopentadienyl ring conferring conformation diversity, and potential antioxidant capacity as well as its ability to generate reactive oxygen species (ROS) under certain conditions when suitable analogues undergo an activation process.[20-26] In addition, ferrocene and its derivatives have shown low toxicity in a wide range of tests in mammals (e.g. mice, rats, monkeys).[20] Recently,

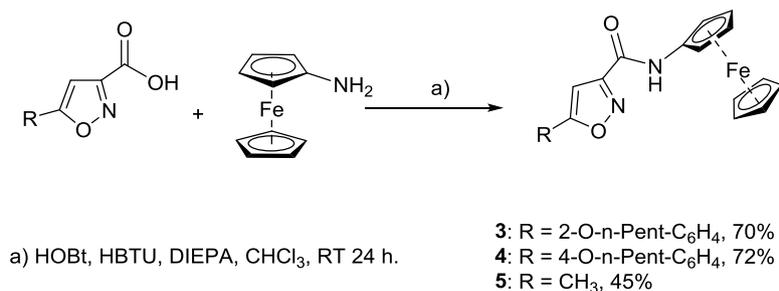
we have performed the replacement of adamantanyl amine by aminoferrocene on a series of in-house dihydroquinoline-based CB<sub>2</sub> ligands. Our study implied that aminoferrocene-based compounds can replace adamantanyl amines in CB<sub>2</sub>-targeting agents without significantly altering the binding affinity and efficacy of the molecules (Figure 2).[27] Given this initial success, we wished to expand this concept to other FAAH and CB<sub>2</sub> –based systems, namely isoxazoles. We anticipated that the formation of direct ferrocenyl analogues of **1** and **2**, namely **3** and **4** (*vide infra*) would suffice in this proof of principle study.



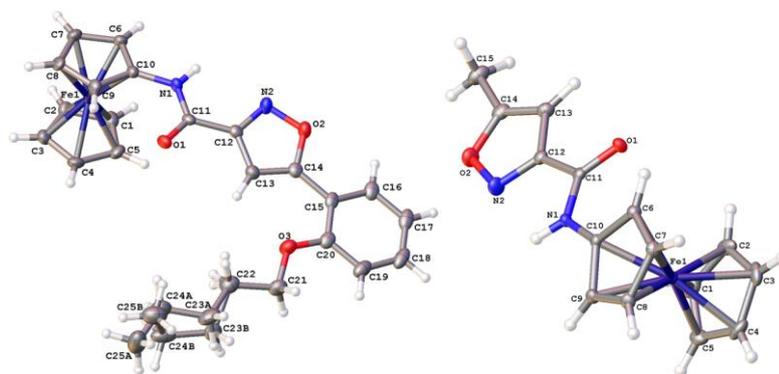
**Figure 2.** The principle of aminoferrocene acting as an adamantyl amine bioisostere.

### Results/Methodology.

Accordingly, we continued this research stream with a view to forming 3-carboxamido-5-ferrocenyl-isoxazole-based modulators of the endocannabinoid system. These compounds were readily made by standard amide coupling protocols and were fully characterized by proton and carbon NMR spectroscopy, mass spectrometry and by elemental analysis. Analogue **5** was also made, but not tested, as it was an excellent candidate for obtaining solid state crystallization data, proving the presence and connectivity of the ferrocene moiety (Figure 3).



**Scheme 1.** Synthesis of isoxazole amides bearing a ferrocenyl unit.



**Figure 3.** Solid state structures of analogues **3** and **5**.

The corresponding derivatives **3** and **4** (Scheme 1) were tested for their  $\text{CB}_2$  vs  $\text{CB}_1$  affinity in a binding assay, their efficacy toward  $\text{CB}_2$ , and their ability to inhibit FAAH.

The affinities of each synthesized compound for both  $\text{CB}_1$  and  $\text{CB}_2$  receptors were determined by a competitive radioligand displacement assay using the dual  $\text{CB}_1/\text{CB}_2$  ligand [ $^3\text{H}$ ]-CP55,940.[16, 28] Compounds displaying potent  $\text{CB}_2$  affinity were further studied for their efficacy by cAMP assays using Chinese hamster ovary cells expressing  $\text{CB}_2$  receptors (CHO- $\text{CB}_2$ ).[29, 30] Cells were treated with forskolin in order to activate adenylyl cyclase-dependent cAMP accumulation. Due to their influence on cAMP formation,  $\text{CB}_2$  ligands can be classified as agonists that promote

cAMP accumulation and inverse agonists that inhibit cAMP production. As illustrated in Table 1, the replacement of the adamantanyl group of compound **1** ( $K_i = 36$  nM) by a ferrocene unit (**3**,  $K_i = 32.5$  nM) did not adversely affect its binding affinity toward CB<sub>2</sub> receptors. Interestingly, such a replacement improved the efficacy of cAMP formation ( $E_{max}$  from 242% to 400%, EC<sub>50</sub> from 1046 nM to 221 nM). This observation is consistent with our previous conclusion that aminoferrocene-based compounds can replace adamantanyl amines in CB<sub>2</sub>-targeting agents.[27] On the contrary, the introduction of a ferrocene unit to replace the adamantanyl group of compound **2** brought about a complete loss of FAAH inhibition probably due to the steric interaction which is induced by the slightly larger ferrocene unit rather than the adamantanyl group. Piomell and co-workers has studied the structure-activity relationship of carbamate-based FAAH inhibitors, which indicated the bulky groups (e.g., *exo*-2-norbornyl, adamantanyl) were unfavorable for FAAH inhibition.[31] Therefore, a ferrocene unit may be more sensitive to the steric interaction with FAAH active site in comparison to an adamantanyl group.

**Table 1.** Affinities ( $K_i$  values), maximum efficacy ( $E_{max}$ ), and/or half-maximal response (EC<sub>50</sub>) toward *h*CB<sub>2</sub> and *h*CB<sub>1</sub> cannabinoid receptors and FAAH inhibition of compounds **1-4**. Selectivity ratios *h*CB<sub>2</sub> versus *h*CB<sub>1</sub>, and cytotoxicity on CHO-WT, CHO-CB<sub>2</sub>, and HT29 Cells.

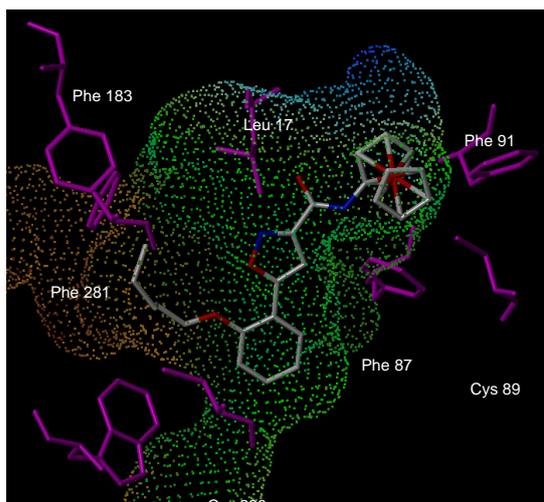
Cpds	<i>h</i> CB <sub>2</sub> and <i>h</i> CB <sub>1</sub> binding assays <sup>a</sup>			CB <sub>2</sub> cAMP assay <sup>a</sup>		FAAH inhibition	Cytotoxicity assays % inhibition at 10 μM		
	CB <sub>2</sub> $K_i$ (nM)	CB <sub>1</sub> $K_i$ (nM)	Ratio CB <sub>1</sub> /CB <sub>2</sub>	$E_{max}$ (%) <sup>b</sup>	EC <sub>50</sub> (nM)	IC <sub>50</sub> (nM)	CHO-WT	CHO-CB <sub>2</sub>	HT29
<b>1</b>	36.0 ± 3.4 <sup>c</sup>	> 1000 <sup>c</sup>	> 83	242 ± 42	1046 ± 400	> 10000 <sup>c</sup>	0%	0%	0 %
<b>2</b>	> 1000 <sup>c</sup>	N. D. <sup>d</sup>	N. D. <sup>d</sup>	N. D. <sup>d</sup>	N. D. <sup>d</sup>	8 <sup>c</sup>	0%	0%	0%
<b>3</b>	32.5 ± 11.4	2513.3 ± 731.6	77.3	400 ± 20	221 ± 52	> 10000	0%	0%	0%
<b>4</b>	> 1000	N. D. <sup>d</sup>	N. D. <sup>d</sup>	N. D. <sup>d</sup>	N. D. <sup>d</sup>	> 10000	1%	7%	10%

WIN55,21					
2	6.9 ± 2.0	13.9 ± 4.0	2.0	45 ± 7	4.3 ± 1.1
(agonist)					
AM630					
(inverse	31.2 ±	5152 ±	165 <sup>e</sup>	232 ± 79	785 ± 7
agonist)	12.4 <sup>e</sup>	567 <sup>e</sup>			

<sup>a</sup>Data represent the mean ± SEM of three or four experiments performed in duplicate or triplicate. <sup>b</sup>E<sub>max</sub> values are expressed as a percentage of forskolin-induced cAMP production. <sup>c</sup>Data from ref 18. <sup>d</sup>N.D. means not determined.

<sup>e</sup>Data from ref 9.

A model of CB<sub>2</sub> was built by homology with the crystallographically resolved complex of CB<sub>1</sub> with the CB<sub>1</sub> taranabant ligand (5u09) [32] in order to study the binding mode of compound **3**. The docking of compound **3** was realized using GOLD 5.1.[33] Compound **3** binds in a V shaped conformation, with a rather large number of hydrophobic contacts, as befits for a cannabinoid receptor (Figure 4). The ferrocene moiety is accommodated by a wide cavity at the extracellular end of the pocket and its pentyl tail points toward a narrower subpocket. Interestingly, the whole molecule is more remote from TM7 than from the other six helices. It does not form any hydrogen bond with the receptor, which is compensated for by the large number of favourable hydrophobic contacts and the orientation of the more hydrophilic parts toward the middle of the cavity. The ferrocene appears to be a key element in the positioning of the compound as it can only fit in the upper part of the cavity due to its width and therefore orients the whole binding mode of the molecule.



**Figure 4:** Putative binding mode of compound **3** into the CB<sub>2</sub> receptor

## Experimental.

### Chemistry.

Aminoferrocene was purchased from TCI, UK and used as such. The two isoxazole acid precursors (i.e., 5-(2-pentyloxyphenyl)isoxazole-3-carboxylic acid and 5-(4-pentyloxyphenyl)isoxazole-3-carboxylic acid) were obtained using known synthetic procedures [16]. High resolution mass spectrometry (HRMS) was performed by the EPSRC National Mass Spectrometry Facility, University of Swansea. Elemental analyses were conducted by Stephen Boyer (London Metropolitan University). Reactions were carried out in a well vented fume hood. <sup>1</sup>H and <sup>13</sup>C-NMR spectra were recorded on Varian 500 MHz or 400 MHz spectrometers and chemical shifts are reported in ppm, usually referenced to TMS as an internal standard.

5-(2-(Pentyloxy)phenyl)-*N*-ferrocenylisoxazole-3-carboxamide, **3**.

5-(2-(Pentyloxy)phenyl)isoxazole-3-carboxylic acid (27.5 mg, 0.1 mmol) was combined with HOBt (6.8 mg, 0.05 mmol), HBTU (57 mg, 0.15 mmol) and DIPEA (0.035 mL) in CHCl<sub>3</sub> (2 mL). The reaction mixture was stirred at room temperature for 45 min and then aminoferrocene (24.1 mg, 0.12 mmol) was added and the mixture was stirred for 24 hr. Thereafter the reaction mixture was washed successively with

NaOH (0.5 N, 20 mL), HCl (1 N, 20 mL) then H<sub>2</sub>O (20 mL). The organic layer was dried with MgSO<sub>4</sub> and evaporated to a ca. 3 mL volume and purified by column chromatography. The orange band was eluted with a hexane: ethyl acetate (1:1) mixture, which was collected and evaporated to dryness. The yield was 34 mg, 70% of an orange solid. Crystallization by diffusion between CH<sub>2</sub>Cl<sub>2</sub> and hexane provided the orange crystals.

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz): δ = 8.00-7.80 (2H, m, Ar), 7.44-7.39 (1H, m, Ar), 7.27 (1H, s, Ar), 7.07-6.93 (2H, m, Ar), 4.76 (2H, d, J = 1.9, 2CH), 4.23 (5H, s, Cp), 4.15 (2H, d, J = 1.9, 2CH), 4.09-3.92 (2H, m, CH<sub>2</sub>), 1.96-1.91 (2H, m, CH<sub>2</sub>), 1.48-1.42 (4H, m, 2CH<sub>2</sub>), 0.97 (3H, t, J = 7.1, CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz): δ = 169.5, 165.2, 159.1, 156.0, 139.8, 131.8, 127.7, 120.6, 112.2, 110.0, 102.8, 69.4, 68.7, 64.9, 61.7, 28.8, 28.2, 22.4, 14.0. HRMS-ESI(m/z): found 481.1184, calcd. for [C<sub>25</sub>H<sub>26</sub>FeN<sub>2</sub>O<sub>3</sub> + Na]<sup>+</sup> 481.1185. Anal. Calcd (%) for C<sub>25</sub>H<sub>26</sub>FeN<sub>2</sub>O<sub>3</sub>: C, 65.51; H, 5.72; N, 6.11. Found (%): C, 65.41; H, 5.80; N, 6.12.

#### 5-(4-(Pentyloxy)phenyl)-*N*-ferrocenylisoxazole-3-carboxamide, **4**.

The title compound was prepared by a coupling reaction. 5-(4-(Pentyloxy)phenyl)isoxazole-3-carboxylic acid (27.5 mg, 0.1 mmol) was reacted with HOBt (6.8 mg, 0.05 mmol), HBTU (57 mg, 0.15 mmol) and DIPEA (0.035 mL) in CH<sub>2</sub>Cl<sub>2</sub> (2 mL). The reaction mixture was stirred at RT for 45 min. and then added aminoferrocene (24.1 mg, 0.12 mmol) stirred for 24 hr. Workup was as above. The orange band was eluted with a hexane: ethyl acetate (1:1) mixture, which was collected and evaporated to dryness. The yield was 35 mg, 72% (orange solid).

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz): δ = 7.94 (1H, s, Ar), 7.75-7.70 (2H, m, Ar), 7.27 (2H, s, Ar), 7.00-6.93 (2H, m, 2CH<sub>2</sub>), 6.89 (1H, s, Ar), 4.75-4.71 (2H, m, 2CH), 4.22 (5H, s, Cp), 4.06-4.01 (2H, m, 2CH), 1.83-1.76 (2H, m, CH<sub>2</sub>), 1.45-1.42 (4H, m, 2CH<sub>2</sub>), 0.96 (3H, s, CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz): δ = 172.0, 161.2, 159.1, 156.9, 127.6, 119.2, 115.1, 97.5, 93.2, 69.3, 68.3, 64.9, 61.7, 28.8, 28.1, 22.4, 14.0. HRMS-ESI(m/z): found 459.1333, calc. for [C<sub>25</sub>H<sub>26</sub>FeN<sub>2</sub>O<sub>3</sub> + H]<sup>+</sup> 459.1366. Anal. Calcd (%) for C<sub>25</sub>H<sub>26</sub>FeN<sub>2</sub>O<sub>3</sub>: C, 65.51; H, 5.72; N, 6.11. Found (%): C, 65.51; H, 5.82; N, 6.18.

### 5-Methyl-*N*-ferrocenylisoxazole-3-carboxamide, **5**.

The title compound was prepared by a coupling reaction. 5-Methylisoxazole-3-carboxylic acid (127 mg, 1 mmol) was reacted with HOBt (68 mg, 0.5 mmol), HBTU (570 mg, 1.5 mmol) and DIPEA (0.35 mL) in CHCl<sub>3</sub> (20 mL). The reaction mixture was stirred at RT for 45 min. and then aminoferrocene (241 mg, 1.2 mmol) was added and the mixture was stirred for 24 hr. Work up was as above. An orange band was eluted with a hexane: ethyl acetate (6:4) mixture, which was collected and evaporated to dryness. The yield was 160 mg, 45% of an orange solid. Crystallization by diffusion between CH<sub>2</sub>Cl<sub>2</sub> and hexane provided orange crystals.

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 500 MHz): δ = 7.90 (1H, s, NH), 6.50 (1H, s, Ar), 4.72-4.69 (2H, m, 2CH), 4.20 (5H, s, Cp), 4.07-3.97 (2H, m, 2CH), 2.52 (3H, s, CH<sub>3</sub>). <sup>13</sup>C NMR (CDCl<sub>3</sub>, 126 MHz): δ = 171.5, 158.8, 156.9, 101.4, 93.3, 76.7, 69.3, 64.9, 61.6, 12.4. HRMS-ESI(m/z): found 311.0471, calcd. for [C<sub>15</sub>H<sub>14</sub>FeN<sub>2</sub>O<sub>2</sub> + H]<sup>+</sup> 311.0477. Anal. Calcd (%) for C<sub>15</sub>H<sub>14</sub>FeN<sub>2</sub>O<sub>2</sub>: C, 58.09; H, 4.55; N, 9.03. Found (%): C, 58.20; H, 4.63; N, 9.13.

### **Biology.**

#### Competition Binding Assay

Stock solutions of the compounds were prepared in DMSO at 10<sup>-2</sup> M and further diluted with the binding buffer to the desired concentration, with a maximal DMSO concentration of 0.1%. Briefly, [<sup>3</sup>H]-CP-55,940 (0.5 nM), nonselective human CB<sub>1</sub> and CB<sub>2</sub> cannabinoid receptor, were added to 6 μg of membranes from CB<sub>1</sub>- or CB<sub>2</sub>-overexpressing CHO cells in binding buffer (50 mM Tris-HCl, 5 mM MgCl<sub>2</sub>, 2.5 mM EDTA, 0.5 mg/mL BSA, pH 7.4). After 90 min at 30 °C, the incubation was stopped and the solutions were rapidly filtered over a UniFilter-96 GF/C glass fiber plate, presoaked in PEI (0.05%) on a Filtermate UniFilter 96-Harvester (PerkinElmer), and washed 10 times with icecold 50 mM Tris-HCl pH 7.4. The radioactivity on the

filters was measured using a TopCount NXT microplate scintillation counter (PerkinElmer) using 30  $\mu$ L of MicroScint 40 (PerkinElmer). Assays were performed at least in triplicate in three independent experiments. The nonspecific binding was determined in the presence of 5  $\mu$ M (R)- (+)-WIN 55,212-2 (Sigma).

### **Conclusion and future perspectives.**

There is growing evidence that CB<sub>2</sub> inverse agonists can attenuate inflammation and osteoporosis through regulating the migration of immune cells.[9-13] Herein, we have shown, albeit limited to 2 examples, that the replacement of adamantylamine by an aminoferrocene is possible and may provide a new perspective for the design of potent CB<sub>2</sub> inverse agonists. This should be readily expandable to other ligand scaffolds other than isoxazoles and dihydroquinilones and to other metallocene-based derivatives. It is anticipated that other GPCR-based or enzyme inhibitor ligands might be designed incorporating a ferrocenyl moiety [34] with the potential for example to assist in X-ray protein studies [35] or to enable further Fe(II)/(III) redox chemistry [36-39].

### **Executive Summary**

- It is possible to synthesize ferrocene analogues of known cannabinoid receptor ligands.
- These are characterized in both solution and in the solid state.
- Compound **3** retains binding and displays improved efficacy at the CB<sub>2</sub> receptor.

- Ongoing studies are looking at other uses of ferrocenes as bioisosteres in bioorganometallic chemistry.

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### **Financial & competing interests disclosure**

The authors have no other relevant affiliations or financial involvement with any organization or entity with a financial interest in or financial conflict with the subject matter or materials discussed in the manuscript apart from those disclosed. No writing assistance was utilized in the production of this manuscript.

**Disclosure.** The work has been previously presented as part of a thesis, see <http://sro.sussex.ac.uk/68599/>.

### **References**

1. Devane WA, Hanus L, Breuer A, Pertwee RG, Stevenson LA, Griffin G, Gibson D, Mandelbaum A, Etinger A, Mechoulam R. Isolation and structure of a brain constituent that binds to the cannabinoid receptor. *Science* 258(5090), 1946-1949 (1992).
2. Simon GM, Cravatt BF. Anandamide biosynthesis catalyzed by the phosphodiesterase gdel and detection of glycerophospho-n-acyl ethanolamine precursors in mouse brain. *J. Biol. Chem.* 283(14), 9341-9349 (2008).
3. Izzo AA, Deutsch DG. Unique pathway for anandamide synthesis and liver regeneration. *Proc. Natl. Acad. Sci. U. S. A.* 108(16), 6339-6340 (2011).
4. Tuo W, Leleu-Chavain N, Spencer J, Sansook S, Millet R, Chavatte P. Therapeutic potential of fatty acid amide hydrolase, monoacylglycerol lipase, and n-acylethanolamine acid amidase inhibitors. *J. Med. Chem.* 60(1), 4-46 (2017).
5. Deutsch DG, Chin SA. Enzymatic synthesis and degradation of anandamide, a cannabinoid receptor agonist. *Biochem. Pharmacol.* 46(5), 791-796 (1993).
6. Piomelli D, Sasso O. Peripheral gating of pain signals by endogenous lipid mediators. *Nat. Neurosci.* 17(2), 164-174 (2014).
7. Beltramo M, Stella N, Calignano A, Lin SY, Makriyannis A, Piomelli D. Functional role of high-affinity anandamide transport, as revealed by selective inhibition. *Science* 277(5329),

- 1094-1097 (1997).
8. Moore SA, Nomikos GG, Dickason-Chesterfield AK, Schober DA, Schaus JM, Ying BP, Xu YC, Phebus L, Simmons RM, Li D, Iyengar S, Felder CC. Identification of a high-affinity binding site involved in the transport of endocannabinoids. *Proc. Natl. Acad. Sci. U. S. A.* 102(49), 17852-17857 (2005).
  9. Ross RA, Brockie HC, Stevenson LA, Murphy VL, Templeton F, Makriyannis A, Pertwee RG. Agonist-inverse agonist characterization at CB<sub>1</sub> and CB<sub>2</sub> cannabinoid receptors of L759633, L759656 and AM630. *Br. J. Pharmacol* 126(3), 665-672 (1999).
  10. Geng D, Xu YZ, Yang HL, Zhu XS, Zhu GM, Wang XB. Inhibition of titanium particle-induced inflammatory osteolysis through inactivation of cannabinoid receptor 2 by AM630. *J. Biomed. Mater. Res. A* 95(1), 321-326 (2010).
  11. Pertwee R, Griffin G, Fernando S, Li X, Hill A, Makriyannis A. AM630, a competitive cannabinoid receptor antagonist. *Life sci.* 56(23-24), 1949-1955 (1995).
  12. Lunn CA, Reich EP, Fine JS, Lavey B, Kozlowski JA, Hipkin RW, Lundell DJ, Bober L. Biology and therapeutic potential of cannabinoid CB<sub>2</sub> receptor inverse agonists. *Br. J. Pharmacol.* 153(2), 226-239 (2008).
  13. Lunn CA, Fine JS, Rojas-Triana A, Jackson JV, Fan X, Kung TT, Gonsiorek W, Schwarz MA, Lavey B, Kozlowski JA, Narula SK, Lundell DJ, Hipkin RW, Bober LA. A novel cannabinoid peripheral cannabinoid receptor-selective inverse agonist blocks leukocyte recruitment in vivo. *J. Pharmacol. Exp. Ther.* 316(2), 780-788 (2006).
  14. Clinical information can be found at <https://www.Clinicaltrials.Gov/>. Drug name: GW842166.
  15. Aghazadeh Tabrizi M, Baraldi PG, Borea PA, Varani K. Medicinal chemistry, pharmacology, and potential therapeutic benefits of cannabinoid CB<sub>2</sub> receptor agonists. *Chem. Rev.* 116(2), 519-560 (2016).
  16. Tourteau A, Andrzejak V, Body-Malapel M, Lemaire L, Lemoine A, Mansouri R, Djouina M, Renault N, El Bakali J, Desreumaux P, Muccioli GG, Lambert DM, Chavatte P, Rigo B, Leleu-Chavain N, Millet R. 3-Carboxamido-5-aryl-isoxazoles as new CB<sub>2</sub> agonists for the treatment of colitis. *Bioorg. Med. Chem.* 21(17), 5383-5394 (2013).
  17. Andrzejak V, Muccioli GG, Body-Malapel M, El Bakali J, Djouina M, Renault N, Chavatte P, Desreumaux P, Lambert DM, Millet R. New FAAH inhibitors based on 3-carboxamido-5-aryl-isoxazole scaffold that protect against experimental colitis. *Bioorg. Med. Chem.* 19(12), 3777-3786 (2011).
  18. Tourteau A, Leleu-Chavain N, Body-Malapel M, Andrzejak V, Barczyk A, Djouina M, Rigo B, Desreumaux P, Chavatte P, Millet R. Switching cannabinoid response from CB<sub>2</sub> agonists to faah inhibitors. *Bioorg. Med. Chem. Lett.* 24(5), 1322-1326 (2014).
  19. Tuo W, Leleu-Chavain N, Barczyk A, Renault N, Lemaire L, Chavatte P, Millet R. Design, synthesis and biological evaluation of potent FAAH inhibitors. *Bioorg. Med. Chem. Lett.* 26(11), 2701-2705 (2016).
  20. Gielen M, Tiekink ER. Metallotherapeutic drugs and metal-based diagnostic agents: the use of metals in medicine. John Wiley & Sons, (2005).
  21. Gasser G, Ott I, Metzler-Nolte N. Organometallic anticancer compounds. *J. Med. Chem.* 54(1), 3-25 (2010).
  22. Yong J, Lu C, Wu X. Synthesis of isoxazole moiety containing ferrocene derivatives and preliminarily in vitro anticancer activity. *MedChemComm* 5(7), 968-972 (2014).

23. Xi GL, Liu ZQ. Solvent-free povarov reaction for synthesizing ferrocenyl quinolines: antioxidant abilities deriving from ferrocene moiety. *Eur. J Med. Chem.* 86, 759-768 (2014).
24. Liu ZQ. Potential applications of ferrocene as a structural feature in antioxidants. *Mini-Rev. Med. Chem.* 11(4), 345-358 (2011).
25. Wang R, Liu ZQ. Ferrocene as a functional group enhances the inhibitive effect of dihydropyrimidine on radical-induced oxidation of DNA. *Org. Chem. Front.* 1(7), 792-797 (2014).
26. Arambula JF, Mccall R, Sidoran KJ, Magda D, Mitchell NA, Bielawski CW, Lynch VM, Sessler JL, Arumugam K. Targeting antioxidant pathways with ferrocenylated N-heterocyclic carbene supported gold (I) complexes in A549 lung cancer cells. *Chem. Sci.* 7(2), 1245-1256 (2016).
27. Sansook S, Tuo W, Lemaire L, Tourteau A, Barczyk A, Dezitter X, Klupsch F, Leleu-Chavain N, Tizzard GJ, Coles SJ, Millet R, Spencer J. Synthesis of bioorganometallic nanomolar-potent CB<sub>2</sub> agonists containing a ferrocene unit. *Organometallics* 35(19), 3361-3368 (2016).
28. Govaerts SJ, Hermans E, Lambert DM. Comparison of cannabinoid ligands affinities and efficacies in murine tissues and in transfected cells expressing human recombinant cannabinoid receptors. *Eur. J. Pharm. Sci.* 23(3), 233-243 (2004).
29. Horváth B, Magid L, Mukhopadhyay P, Bátkai S, Rajesh M, Park O, Tanchian G, Gao RY, Goodfellow CE, Glass M, Mechoulam R, Pacher P. A new cannabinoid CB<sub>2</sub> receptor agonist GU-910 attenuates oxidative stress, inflammation and cell death associated with hepatic ischaemia/reperfusion injury. *Br. J. Pharmacol.* 165(8), 2462-2478 (2012).
30. Marini P, Cascio MG, Pertwee RG. The cyclic amp assay using human cannabinoid CB<sub>2</sub> receptor-transfected cells. *Methods Mol. Biol.* 1412, 85-93 (2016).
31. Mor M, Lodola A, Rivara S, Vacondio F, Duranti A, Tontini A, Sanchini S, Piersanti G, Clapper JR, King AR, Tarzia G, Piomelli D. Synthesis and quantitative structure-activity relationship of fatty acid amide hydrolase inhibitors: modulation at the N-portion of biphenyl-3-yl alkylcarbamates. *J. Med. Chem.* 51 (12), 3487-3498 (2008).
32. Shao Z, Yin J, Chapman K, Grzemska M, Clark L, Wang J, Rosenbaum DM. High-resolution crystal structure of the human CB1 cannabinoid receptor. *Nature* 540, 602-606 (2016).
33. Jones G, Willett P, Glen RC, Leach AR, Taylor R. Development and validation of a genetic algorithm for flexible docking. *J. Mol. Biol.* 267 (3), 727-748 (1997).
34. Maryanoff, BE, Keeley, SL, Persico, FJ. Replacement of aromatic or heteroaromatic groups in nonsteroidal antiinflammatory agents with the ferrocene group. *J. Med. Chem.* 26 (2), 226-229 (1983).
35. Salmon, A.J., Williams, M., Hofmann, A., Poulsen, S.-A. Protein crystal structures with ferrocene and ruthenocene-based enzyme inhibitors. *Chem. Commun.* 48, 2328-2330 (2012).
36. Patra, M, Gasser, G. The medicinal chemistry of ferrocene and its derivatives. *Nat. Rev. Chem.* 1, 0066 (2017).
37. Ocasio, CA Sansook, S, Jones, R, Roberts, J M, Scott, T G, Tsoureas, N, Coxhead, P, Guille, M, Tizzard, G J, Coles, S J, Hocheegger, H, Bradner J E, Spencer, J Pojamide: an HDAC-3-selective ferrocene analogue with remarkably enhanced redox-triggered ferrocenium activity in cells *Organometallics*, 36, 3276-3283 (2017).

38. Jaouen, G, Vessières, A, Top, S. Ferrocifen type anti cancer drugs. *Chem. Soc. Rev.*, **44**, 8802-8817 (2015).

39. Hagen, H, Marzenell, P, Jentsch, E, Wenz, F, Veldwijk, M R, Mokhir, A. Aminoferrocene-Based Prodrugs Activated by Reactive Oxygen Species, *J Med Chem*, *55*(2), 924-934 (2012).