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Disseminated neoplasia in blue mussels, *Mytilus galloprovincialis*, from the Black Sea, Romania

Corina Ciocan¹,* Inke Sunila²

¹National Institute for Marine Research and Development, Grigore Antipa, Mamaia Blvd. 300, Constanta 8700, Romania

²State of Connecticut, Department of Agriculture, Bureau of Aquaculture, PO Box 97, Milford, Connecticut 06460, USA

*Email: C.M.Ciocan@sussex.ac.uk

Fax: 0040 1273 677196

Present address: University of Sussex, Department of Biology and Environmental Sciences, Falmer, Brighton BN1 9RH, UK

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ABSTRACT

Disseminated neoplasia, also called leukemia or hemic neoplasia, has been detected in 15 species of marine bivalve mollusks worldwide. The disease is characterized by the presence of single anaplastic cells with enlarged nuclei and sometimes frequent mitosis, in hemolymph vessels and sinuses. The neoplastic cells gradually replace normal hemocytes leading to the increased mortality of animals. The neoplasia reaches epizootic prevalences in blue mussels, *Mytilus trossulus*, in some areas, whereas prevalences in *M. edulis* are generally very low. *M. galloprovincialis* was suggested to be resistant to the disease although very low prevalences were documented from Spain in the Atlantic Ocean and Italy in the Mediterranean Sea. A case of disseminated neoplasia was discovered in *M. galloprovincialis* from among 200 specimens studied from the coast of the Romanian Black Sea. Histological preparation revealed the presence of large anaplastic cells with lobed nuclei. This observation extends the geographic range of marine bivalve mollusks with disseminated neoplasia to include the Black Sea.
INTRODUCTION

Disseminated neoplasia, also called leukemia or hemic neoplasia, has been reported in 15 species of marine or estuarine bivalve mollusks from around the world. Bivalve families affected with the disease include mussels (Mytilus spp.), oysters (Crassostrea and Ostrea spp.), soft shell clams (Mya spp.), cockles (Cerastoderma spp.) and macoma clams (Macoma spp.) (Peters 1988). The condition is characterized by proliferation of anaplastic circulating cells with relatively large nuclei, a high nucleus-to-cytoplasm ratio and frequent presence of mitosis (Elston et al. 1992). The condition has also been called sarcomatoid proliferative disease (Farley 1969), proliferative atypical hemocytic condition (Lowe and Moore 1978), epizootic sarcoma (Farley et al. 1986), sarcomatous neoplasia (Brousseau 1987), transmissible sarcoma (Farley et al. 1991) and systemic neoplasia (Moore et al. 1991). Regardless the nomenclature, this disorder, apparently of hemocytic origin, is distinguishable from another group of bivalve neoplasia, germinomas (reviewed by Peters et al., 1994) even when these cases become invasive and systemic. Branchial carcinoma of the macoma clam (Farley, 1976) presents some difficulties in distinguishing it from disseminated neoplasia.

Disseminated neoplasia in bivalves is progressive and fatal. Anaplastic cells proliferate and replace normal hemocytes in circulation. Elston et al. (1988a) examined the pathogenesis of the disease in individual mussels (M. trossulus) from hemolymph preparations over a 4-month period. Fifty percent of the mussels progressed to advanced disease, most of which died, and 20% showed early advancement of the disease, but went
into remission. Proliferating anaplastic cells consist of different cell populations with different ploidy levels, penta- and tetraploid, which are reflected as different morphologies (Elston et al. 1990, Moore et al. 1991). Diseased mussels have impaired defense mechanisms after the decline in the number of circulating hemocytes, as demonstrated experimentally by the inability of diseased mussels to clear injected bacteria (Kent et al. 1989).

This disease can be transmitted by inoculating neoplastic cells into healthy mussels (Elston et al. 1988b). Whether the disease in these experiments was transmitted via cell transplantation or an infective agent was unclear. In another bivalve, the soft clam, evidence of retroviral origin was reported in three separate studies (Oprandy et al. 1981, 1983, Medina et al. 1993, House et al. 1998). Interestingly, attempts to transmit mussel neoplasia to other bivalve species such as soft clams (Mya arenaria), flat oysters (Ostrea edulis) and Olympia oysters (Ostrea conchaphila), failed. Only mussels developed the disease (Kent et al. 1991).

In some areas disseminated neoplasia reaches epizootic prevalence in selected bivalve species causing serious regional economic damage to the aquaculture industry. Up to 40% prevalence was reported in Mytilus trossulus in Puget Sound, Washington, USA (Elston et al. 1988a), 72% in Cerastoderma edule in Cork Harbor, Ireland (Twomey & Mulcahy 1988) and 90% in Mya arenaria in New Bedford Harbor, Massachusetts, USA (Reinisch et al. 1984). The susceptibility to the disease differs in different species/subspecies of Mytilus. A single case of neoplasia in Mytilus galloprovincialis was reported among several thousand mussels studied for the Mussel Watch program in
California, USA (Hillman 1990). During a pathology survey of mussels (*Mytilus galloprovincialis*) farmed in Taranto, Manfredonia and Cagnano Varano (Italy) carried out between February and December of 1990, neoplasia was found in two Taranto mussels sampled in March and in four others sampled in July (Tiscar et al. 1990, Zizzo et al. 1991). Another single case of “haemocytic neoplasia” was described in cultured mussels *Mytilus galloprovincialis* from Ria de Arousa, NW of Spain (Figueras et al. 1991a). More than 10 years later, Fuentes et al. (2002) reported disseminated neoplasia in five (out of 135) hatchery-reared mussels from the same location. According to Villalba et al. (1997) disseminated neoplasia should not be considered as a threat to the mussel culture industry in Galicia (NW of Spain) since its prevalence is very low in the region: only five mussels (*Mytilus galloprovincialis*) with a cellular proliferative disorder were reported in 1997. Nevertheless, Elston et al. (1992) suggested that *M. galloprovincialis* is resistant to leukemia.

Here, we report a single case of disseminated neoplasia in *Mytilus galloprovincialis* from Romania, the Black Sea. This is the first report of bivalve neoplasia in the region.

**MATERIALS AND METHODS**

Mature wild mussels, *Mytilus galloprovincialis*, were collected by divers offshore near Navodari resort, 5 km north of Constanta, Romanian Black Sea coast, during April 1996. 200 individuals of *Mytilus* (50-70mm length) were sampled from a depth of 2.0-2.5m at 15 days intervals (100 individuals/batch).
At the laboratory, mussels were immediately shucked and the tissues preserved in 10% formalin solution for 3-5 days. Specimens were embedded in paraffin and 5µm tissue sections were stained with hematoxylin-eosin-methyl blue (a modified Masson Trichrome). The slides were examined for gonad development to identify the spring spawning peak, but the individuals were also diagnosed for possible histopathological changes as the sampling site is situated close to a small harbor (Midia Harbor).

RESULTS

One of the histological sections contained mainly gill tissue demonstrating transverse sections of gill filaments. It was characterized by the presence of large, atypical, neoplastic cells (Fig. 1A). Branchial vessels contained no normal hemocytes, but were instead filled with round anaplastic cells (7.36µm, SD=0.80, n=50). Cells had eccentric hyperchromatic indented, lobed or otherwise pleomorphic nuclei (Fig. 1B). Chromatin was granular and no nucleoli were observed. Nucleus to cytoplasm ratios was high and only scant, pale-staining cytoplasm was present. Some cells were binucleated, and a metaphase plate was observed.

DISCUSSION

Mussels (*Mytilus* sp.) have been used as model organisms in histopathology and genetic studies more than any other group of marine invertebrates.
The *Mytilus* spp is a diverse group of bivalves with a broad distribution in the marine environment. Much of the confusion in mussel taxonomy has arisen because of the emphasis placed on shell morphology. Such characteristics are enormously plastic, being subjected to a wide range of environmental factors. Prior to the use of electrophoresis, about nine distinct species of *Mytilus* were recognized: *Mytilus edulis* from northern temperate latitudes, *Mytilus galloprovincialis* from the Mediterranean Sea, *Mytilus trossulus* from the Pacific coast of North America, *Mytilus coruscus* from Japan and China, *Mytilus californianus* from the Pacific coast of North America, *Mytilus chilensis* from Chile, *Mytilus platensis* from Argentina, *Mytilus planulatus* from Australia and *Mytilus desolationis* from the Kerguelen Islands in the southern Indian Ocean (Gosling 1992).

Disseminated neoplasia in blue mussels has been reported from numerous locations worldwide in four different *Mytilus* species (Fig. 2). The reports include *Mytilus galloprovincialis* from the Black Sea (1), Romania (the present study), Mediterranean Sea (2), Italy (Tiscar et al. 1990, Zizzo et al. 1991) and East Atlantic Ocean (3), Galicia, Spain (Figueras et al. 1991, Fuentes et al., 2002, Villalba et al. 1997). Further, there was a single case of disseminated neoplasia in the East Pacific (9), in California, USA in *M. galloprovincialis* (Hillman 1990). Neoplasia has been detected in *M. edulis* in the UK in the Irish Sea (4) and the North Sea (5) (Lowe & Moore 1978, Green & Alderman 1983) and in *M. trossulus* in the Baltic Sea (6), Denmark by Rasmussen (1986) and in Finland by Sunila (1987). In North America, in the West Atlantic, Figueras et al. (1991b) reported a case of disseminated neoplasia in *M. edulis* in Maine, USA, and Hillman et al.
(1992) reported seven cases during six years of the Mussel Watch Program among several thousand mussels studied in Massachusetts, Connecticut and New York (7). Disseminated neoplasia in *M. trossulus* from the Northeast Pacific (8) was reported in several articles in British Columbia, Canada (Cosson-Mannevy et al. 1984, Bower 1989), Oregon (Farley 1969, Mix 1983) and Washington State, USA (Elston et al. 1988a). In South America, in the Southeast Pacific, there is a single report of disseminated neoplasia in *M. chilensis* (Campalans et al. 1998) in Chiloe Island, Chile (10). Finally, a case of disseminated neoplasia in *M. trossulus* was reported in the West Pacific, Sea of Japan (11) at Nakhodka Bay close to Vladivostok, Russia (Usheva & Frolova 2000).

Disseminated neoplasia occurs in *Mytilus galloprovincialis* and *Mytilus edulis* at very low prevalences. Interestingly, Fuentes et al. (2002) suggested the existence of a depressed immune system in hybrid mussels (*Mytilus edulis/Mytilus galloprovincialis*), explaining their higher susceptibility to neoplasia compared with mussels from pure crosses. The case of disseminated neoplasia in mussels from the Black Sea presented in this work, in association with similar cases in different *Mytilus* species reviewed above, demonstrates a wide geographical distribution of disseminated neoplasia in mussels in the world’s oceans and estuaries.

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Fig. 1. A. Disseminated neoplasia in the mussel *Mytilus galloprovincialis* from Romania, Black Sea. Gill filaments with neoplastic cells. G=gill filament, arrows point to neoplastic cells. Paraffin section, Masson-Gomori Trichrome stain. Scale bar 50µm. B. Higher magnification of Fig. 1B. Arrows point to neoplastic cells. Scale bar 25µm.
Fig. 2. Geographical distribution of disseminated neoplasia in different species of blue mussels *Mytilus* sp.

1, 2 and 8  *M. galloprovincialis*  
3, 4 and 6  *M. edulis*  
5, 7 and 10 *M. trossulus*  
9  *M. chilensis*