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The relationship between managed bees and the prevalence of parasites in bumblebees

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ABSTRACT

Honey bees and, more recently, bumblebees have been domesticated and are now managed commercially primarily for crop pollination, mixing with wild pollinators during foraging on shared flower resources. There is mounting evidence that managed honey bees or commercially produced bumblebees may affect the health of wild pollinators such as bumblebees by increasing competition for resources and the prevalence of parasites in wild bees. Here we screened 764 bumblebees from around five greenhouses that either used commercially produced bumblebees or did not, as well as bumblebees from 10 colonies placed at two sites either close to or far from a honey bee apiary, for the parasites *Apicystis bombi*, *Crithidia bombi*, *Nosema bombi*, *N. ceranae*, *N. apis* and deformed wing virus. We found that *A. bombi* and *C. bombi* were more prevalent around greenhouses using commercially produced bumblebees, while *C. bombi* was 18% more prevalent in bumblebees at the site near to the honey bee apiary than those at the site far from the apiary. Whilst these results are from only a limited number of sites, they support previous reports of parasite spillover from commercially produced bumblebees to wild bumblebees, and suggest that the impact of stress from competing with managed bees or the vectoring of parasites by them on parasite prevalence in wild bees needs further investigation. It appears increasingly likely that the use of managed bees comes at a cost of increased parasites in wild bumblebees, which is not only a concern for bumblebee conservation, but which may impact other pollinators as well.

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INTRODUCTION

In recent years several bumblebee species as well as other pollinators have suffered range declines in parts of Europe, the Americas and Asia (*Biesmeijer et al., 2006; Cameron et al., 2011; Goulson, Lye & Darvill, 2008; Potts et al., 2010*). Changes in anthropogenic land-use is a major contributing factor to these declines, with agricultural intensification reducing floral diversity and nesting habitats from many pollinators (*Goulson et al., 2005; Ricketts et al., 2008; Vanbergen et al., 2013*). This has left some bumblebee species fragmented, in small populations with low genetic diversity, something which may make bees more vulnerable

to stresses such as parasites ([Darvill et al., 2006](#); [Ellis et al., 2006](#); [Evison et al., 2013](#); [Oldroyd, 2007](#); [Whitehorn et al., 2011](#)).

In addition to the stresses of habitat loss, pesticide exposure and natural parasites, ([Goulson, 2003](#)), the use of managed bees may place additional stresses on bumblebee populations. Honey bees have been managed commercially for crop pollination and honey production for centuries, and are often kept in commercial apiaries with tens to thousands of colonies, substantially increasing the density of bees in an area. Bumblebees are also now commercially produced and used mainly in greenhouses, but also sometimes in polytunnels and open crops, in Europe, North America, South America, New Zealand and Asia to enhance the yields of soft fruit crops ([Velthuis & van Doorn, 2006](#)). Although the greenhouses in which commercially produced bumblebees are most commonly used are meant to be closed, the commercially produced bumblebees are frequently found foraging outside the greenhouses, and wild bees have been found foraging inside them ([Kraus et al., 2011](#); [Morandin et al., 2001](#); [Murray et al., 2013](#); [Whittington et al., 2004](#)). By freely mixing with wild bumblebees, the deployment of commercially produced bumblebees effectively increases the local density of bumblebees. Bumblebee parasites can be dispersed between bumblebees following shared flower usage ([Durrer & Schmid-Hempel, 1994](#)), and, as a result, the rate of parasite transmission between bees will predictably rise with increased pollinator density ([Arneberg et al., 1998](#)). In areas utilising commercially produced bumblebees, higher parasite prevalence may be expected to be the result, due to either the spillover of parasites from the commercially produced bumblebees, parasite spillback from wild bumblebees, or stress related to the high pollinator density ([Kelly et al., 2009](#); [Power & Mitchell, 2004](#); [Schmid-Hempel, 2011](#)).

The spillover of parasites from one host to another, either intraspecifically or interspecifically, is well known for many organisms ([Power & Mitchell, 2004](#)). There is now good evidence that the honey bee parasites *Nosema ceranae* and deformed wing virus have spilled over to bumblebees, with both being virulent and now widespread in their new bumblebee host ([Evison et al., 2012](#); [Furst et al., 2014](#); [Genersch et al., 2006](#); [Graystock et al., 2013a](#); [Plischuk et al., 2009](#)). In addition, parasites may also spill over to wild bumblebees from the commercially reared bumblebees used in greenhouses. Colonies of commercially produced bumblebees have been shown in many studies to carry parasites ([Colla et al., 2006](#); [Gegear, Otterstatter & Thomson, 2005](#); [Manson, Otterstatter & Thomson, 2010](#); [Meeus et al., 2011](#); [Murray et al., 2013](#); [Otterstatter & Thomson, 2007](#); [Singh et al., 2010](#); [Whittington & Winston, 2003](#)), with the most recent study using sensitive molecular methods finding that three-quarters of the colonies investigated were infected by at least one parasite and confirming that these parasites were in many cases infectious ([Graystock et al., 2013b](#)). The introduction of commercially produced bumblebees has been associated with the introduction of foreign parasites and correlated declines in native bumblebee species in Japan, South America and North America, suggesting that the spillover of parasites has occurred on multiple occasions ([Arbetman et al., 2012](#); [Colla et al., 2006](#); [Goka et al., 2001](#); [Meeus et al., 2011](#); [Otterstatter & Thomson, 2008](#); [Szabo et al., 2012](#)).

Although attention has focussed on parasite spillover, it is also possible that the use of managed honey bees and commercially produced bumblebees may increase the prevalence of parasites in wild bumblebees via parasite spillback or heightened stress from increased competition when foraging. Managed honey bees or commercially produced bumblebees may become infected with parasites carried by the wild bees, and their unnaturally high density in apiaries or greenhouses may then result in them acting as a reservoir in which the prevalence of parasites becomes high, from which the parasites can then spillback into wild bees ([Kelly et al., 2009](#)). The increased competition for resources caused by the introduction of high densities of managed honey bees or commercially produced bumblebees may also stress wild bees due to the increased competition when foraging, which can have negative effects on various fitness components including resistance to parasites ([Brown, Loosli & Schmid-Hempel, 2000](#); [Elbgami et al., 2014](#); [Foley et al., 2012](#); [Goulson & Sparrow, 2009](#); [Lafferty & Gerber, 2002](#); [Mallon, Brockmann & Schmid-Hempel, 2003](#)).

The prevalence of parasites in wild bumblebees appears to be greater when the bees are in proximity to greenhouses using commercially produced bumblebee colonies ([Colla et al., 2006](#); [Murray et al., 2013](#); [Otterstatter & Thomson, 2008](#)). However, whether this is due to parasite spillover, parasite spillback, or stress, is not always clear. Here we investigate the relationships between commercially reared bumblebees or managed honey bees and the prevalence of a range of parasites in bumblebees. We first examine the relationship between the prevalence of parasites in wild bumblebees and proximity to three farms in which commercially reared bumblebees being used and two greenhouse farms in which they were not being used. In addition, we examine the effect of proximity to honey bees on bumblebee parasite prevalence, using bumblebee colonies located at two sites, either near or far from an apiary.

MATERIALS AND METHODS

The effect of proximity to commercially reared bumblebees

To determine the prevalence of parasites at sites either using commercially produced bumblebees or not, five greenhouse farm sites in England were selected. Sites were selected based on the presence of large scale commercial fruit farms (ca. 50–75 ha) that utilised greenhouses and/or polytunnels for crop growing. Sites were all of comparable size, located in areas of open farmland with no other sites known to be deploying bumblebees within 10 km. Three of the sites in Cambridgeshire, Kent and Essex, were a focal greenhouse in which commercially produced bumblebees were used for the pollination of the greenhouse crops (≈ 200 –300 hives at each site), and two sites in Merseyside and Oxfordshire were a focal greenhouse in which commercially produced bumblebees had not been used (all sites were at least 70 km apart). Bumblebees were collected with a sweep net within 0.5 km of points 0.5, 3 and 5 km from the focal greenhouse sites, with approximately 50 bumblebees collected at each of the three distances for each of the five sites. All bees were collected over a 16 day period in the summer of 2011 (Cambridgeshire, Kent and Essex on 2nd July, 9th July and 11th July respectively, Oxfordshire and Merseyside on 1st

and 16th July respectively). A total of 471 bumblebees were collected from around the sites using commercially produced bumblebees (222, 151 and 98 at the Cambridgeshire, Kent and Essex sites respectively) and a total of 293 bumblebees from around the sites not using commercially produced bumblebees (143 and 150 at the Merseyside and Oxfordshire sites respectively). The samples consisted of *B. terrestris*, *B. hortorum*, *B. hypnorum*, *B. lapidarius*, *B. lucorum*, *B. pascuorum* and *B. pratorum*, with most being either *B. terrestris* or *B. lapidarius* (40% and 25% of samples respectively; see [Table 1](#) for detail of the number of each species sampled at each distance at each site). All of these 764 bumblebees were screened for parasites.

The effect of proximity to managed honey bees

Ten commercially produced *Bombus terrestris audax* bumblebee colonies (Biobest) with 80–100 workers were used to determine the effect of proximity to managed honey bee colonies on parasite prevalence within bumblebee colonies. The colonies were placed on a farm near Tadcaster, West Yorkshire (53°52'N, 1°20'W). Five of the bumblebee colonies were situated on the edge of an agroforestry field containing an apiary with 50, full-size honey bee hives, and the remaining five bumblebee colonies were sited at the edge of a field 1 km away from the apiary, with bees at both locations being in the same landscape with access to similar floral resources ([Elbgami et al., 2014](#)). The colonies were placed in a row at the edge of each site, with the same distance between hives in each case. The bumblebee colonies remained at these sites for one month, during which they could forage freely. After this period, 20 bumblebee workers were taken from each colony and screened for the presence of the parasites.

Molecular screening for parasite presence

A ca. 0.5 cm³ sample of midgut, malpighian tubules and fatbody from each bee was homogenised and DNA extracted from the homogenate using 5% Chelex. All DNA samples were amplified for the 18S Apidae host control gene to confirm the quality of the DNA extraction. Samples were then screened for the presence of the *Apicystis bombi*, *Crithidia bombi*, *Nosema bombi*, *N. ceranae*, *N. apis* and deformed wing virus (DWV) parasites using parasite specific primers and conditions ([Chen, Higgins & Feldlaufer, 2005](#); [Gisder & Genersch, 2013](#); [Klee, Tek Tay & Paxton, 2006](#); [Meeus et al., 2010](#); [Table S1](#)). Products were run alongside a size standard on a 1% agarose gel stained with ethidium bromide to confirm amplicon size. Each assay included a negative and a positive control.

Statistical analysis

The prevalence and richness of parasites was compared between sites in which greenhouses did or did not use commercially produced bumblebees, and between the sites near to or far from the honey bee apiary using generalized linear models (GLM) with the likelihood ratio χ^2 statistic. The parasite richness (number of parasite species detected in a single host) was compared between sites using a negative binomial distribution and log link function and changes in the prevalence of individual parasites with a binomial distribution and logit link function. When looking at the effect of commercially produced bumblebees, site

Table 1 Summary of the bumblebees surveyed. A variety of wild bumblebees were captured to assess the prevalence of parasites in bumblebees sampled at three distances from greenhouses that either were or were not using commercially produced bumblebees. The species, frequency, sex and location of wild bumblebee collected are shown along with their collection site specifics.

Location	Species	N	Distance from greenhouses ± 0.5 km					
			0.5 km		3 km		5 km	
			F	M	F	M	F	M
Cambridgeshire Latitude: 52°18'0.79"N Longitude: 0°3'2.46"W Area: ≈ 50 acre Number of hives: $\approx 200^a$	<i>B. hortorum</i>	6	3	0	1	0	2	0
	<i>B. hypnorum</i>	9	3	0	3	0	3	0
	<i>B. lapidarius</i>	99	16	4	43	5	25	6
	<i>B. lucorum</i>	33	6	5	4	2	12	4
	<i>B. pascuorum</i>	15	5	0	3	0	6	0
	<i>B. pratorum</i>	16	0	5	5	2	3	2
Kent Latitude: 51°21'13.64"N Longitude: 1°17'8.00"E Area: ≈ 75 acre Number of hives: $\approx 300^a$	<i>B. terrestris</i>	44	15	5	6	0	16	2
	<i>B. lapidarius</i>	21	6	1	5	1	8	0
	<i>B. lucorum</i>	12	4	0	3	1	4	0
	<i>B. pascuorum</i>	5	3	0	2	0	0	0
	<i>B. pratorum</i>	19	8	0	3	1	6	1
	<i>B. terrestris</i>	94	26	3	31	3	28	3
Essex Latitude: 51°56'0.67"N Longitude: 1°0'18.17"E Area: ≈ 60 acre Number of hives: $\approx 240^a$	<i>B. lapidarius</i>	30	6	0	5	9	7	3
	<i>B. lucorum</i>	2	0	0	2	0	0	0
	<i>B. pascuorum</i>	38	16	2	10	1	9	0
	<i>B. pratorum</i>	25	21	2	0	1	0	1
	<i>B. terrestris</i>	3	3	0	0	0	0	0
	<i>B. hypnorum</i>	9	2	0	2	0	5	0
Merseyside Latitude: 53°30'40.61"N Longitude: 2°47'17.78"W Area: ≈ 75 acre	<i>B. lapidarius</i>	17	6	0	2	1	8	0
	<i>B. lucorum</i>	30	5	0	11	1	11	2
	<i>B. pascuorum</i>	2	0	0	0	0	1	1
	<i>B. pratorum</i>	12	6	0	2	0	3	1
	<i>B. terrestris</i>	73	28	3	23	1	15	3
	<i>B. lapidarius</i>	25	9	0	8	1	6	1
Oxfordshire Latitude: 51°40'10.01"N Longitude: 1°22'38.79"W Area: ≈ 50 acre	<i>B. lucorum</i>	9	3	0	2	0	4	0
	<i>B. pascuorum</i>	12	5	0	4	0	3	0
	<i>B. pratorum</i>	16	5	0	2	3	6	0
	<i>B. terrestris</i>	88	27	1	29	1	28	2

Notes.

^a Numbers of hives estimated based on size of farm. As a general rule, producers recommend using 4 bumblebee hives/acre at the beginning of the season, then systematically introducing more hives as the original ones age. The estimates here are based on 4 hives/acre.

type (greenhouses in which commercially produced bumblebees were or were not used), transect distance, and site location nested within site type were included as factors, with the species and sex of bumblebees sampled also included as factors. We did not include sampling dates in these models because it covaried with site, but checked for temporal autocorrelation using Box–Ljung tests and retested the GLM without site and instead including sampling date (number of days after the first sample was collected) as a covariate. We checked for spatial autocorrelation using Moran's I (Rogerson, 2010). When looking

at the effect of managed honey bees, location (near to or far from the apiary), and colony nested within location, were used as factors. Nonsignificant terms were removed stepwise based on log-likelihood ratio tests in all cases to obtain the minimum adequate models (Table S2). All analyses were carried out in PASW Statistics 20 (IBM, Armonk, NY, USA).

RESULTS

The effect of commercially produced bumblebees on parasite prevalence in wild bumblebees

Overall, most wild bumblebees had either no infections (40.7%) or infection by a single parasite species (40.3%), with cases of bumblebees infected by two or three parasite species being rare (16.8% and 2.1% respectively). There was a significant interaction between the distance from the greenhouses and whether they were or were not using commercially produced bumblebees on the numbers of parasite species that infected bumblebees ($\chi^2 = 6.78$, d.f. = 2, $P = 0.034$), and this was not affected by either the species or sex of the bumblebee ($\chi^2 = 3.04$, d.f. = 6, $P = 0.804$ and $\chi^2 = 0.874$, d.f. = 1, $P = 0.35$, respectively). The numbers of parasite species recorded decreased with distance from the greenhouses at sites which were using commercially produced bumblebees, but were unaffected by distance at the sites which were not using these bees (Fig. 1A). There was also a significant difference between individual sites nested within categories of using or not using commercially produced bumblebees ($\chi^2 = 29.0$, d.f. = 3, $P < 0.001$), but the bees sampled from around sites using commercially produced bumblebees nevertheless had significantly more parasite species overall when controlling for this ($\chi^2 = 23.2$, d.f. = 1, $P < 0.001$). The samples were collected within a relatively short 16 day period and showed no temporal autocorrelation (Box–Ljung tests all $P > 0.05$), with there being no effect of sampling date on the number of parasite species found ($\chi^2 = 2.86$, d.f. = 1, $P = 0.091$). There was also no spatial autocorrelation (Moran's $I = 0.062$; a value of 1 indicates perfect correlation and of 0 indicates random dispersion). Of the individual parasites, the prevalence of both *A. bombi* and *N. ceranae* were affected significantly by the interaction between distance and whether sites were using commercially produced bumblebees or not ($\chi^2 = 44.5$, d.f. = 2, $P < 0.001$, and $\chi^2 = 7.98$, d.f. = 2, $P = 0.019$, respectively). *A. bombi* was more commonly close to greenhouses when those greenhouses were using commercially produced bumblebees but showing little effect of distance when they were not (Fig. 1B). *N. ceranae*, in contrast, increased with distance from the greenhouses at the sites not using commercially produced bumblebees but was little affected by distance at the sites where they were (Fig. 1D). *Crithidia bombi* was more prevalent in bumblebees caught from around sites using commercially produced bumblebees than those not using them ($\chi^2 = 15.1$, d.f. = 1, $P < 0.001$) but displayed no proximity effect ($\chi^2 = 0.756$, d.f. = 2, $P = 0.685$; Fig. 1C). *N. ceranae* was the only parasite to show a significant effect of the species or sex of bumblebee sampled (Table S2), which was due to all 7 of the *B. hortorum* bees sampled being workers that were infected by *N. ceranae*. The prevalence of *N. bombi*, *N. apis* and DWV in bumblebees caught were all under 1% and were not affected by any variables (Table S2; Fig. 1).

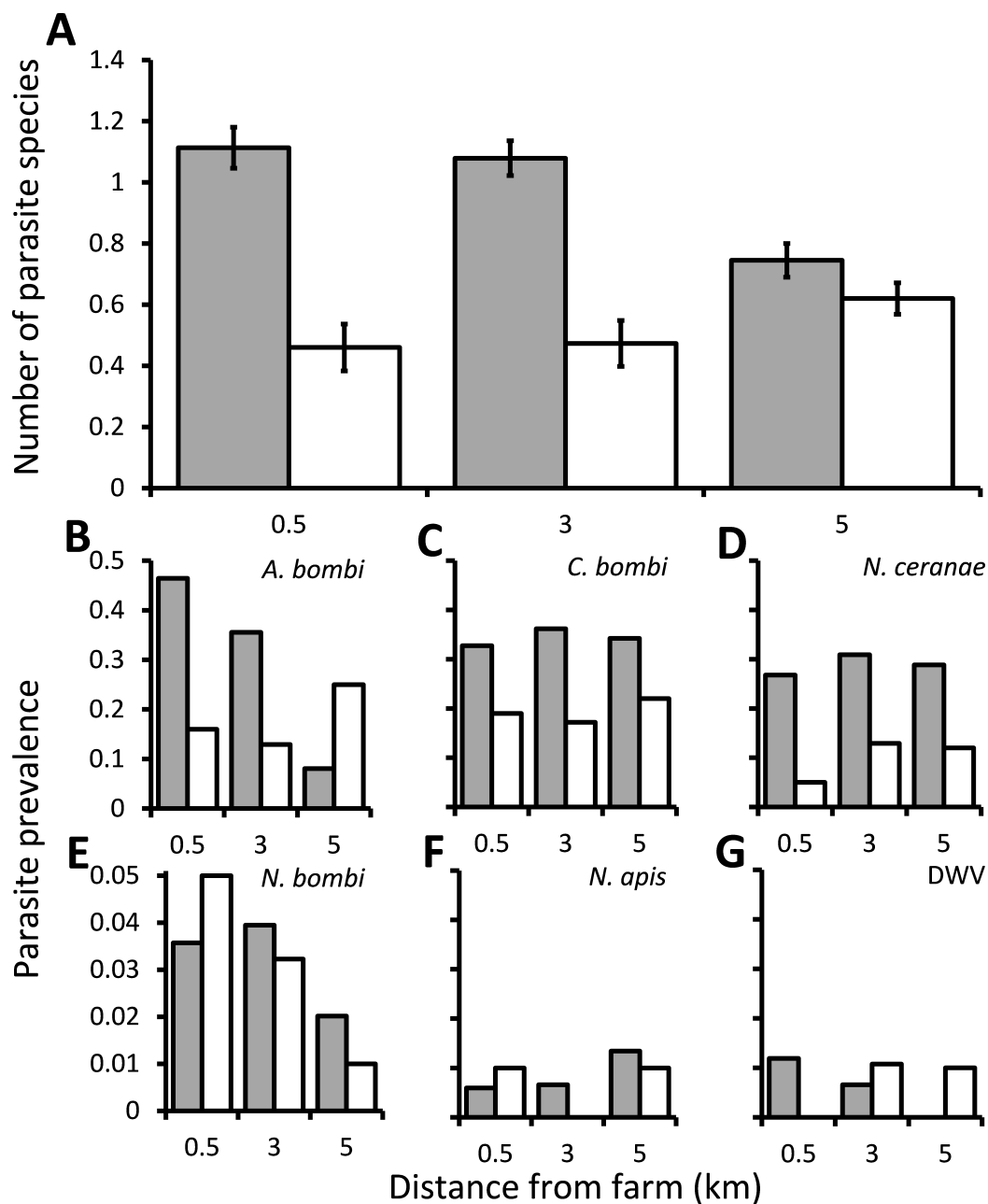


Figure 1 The effect of commercially produced bumblebees on parasite prevalence. Prevalence of parasites in bumblebees sampled within 0.5 km of locations that were 0.5, 3 or 5 km from greenhouses that were either using (grey columns) or not using (white columns) commercially produced bumblebee colonies. (A) The mean \pm s.e. parasite richness (number of species) infecting individual bees. (B–G) The proportion of bumblebees sampled which were positive for the *A. bombi*, *C. bombi*, *N. ceranae*, *N. bombi*, *N. apis* and deformed wing virus (DWV) parasites.

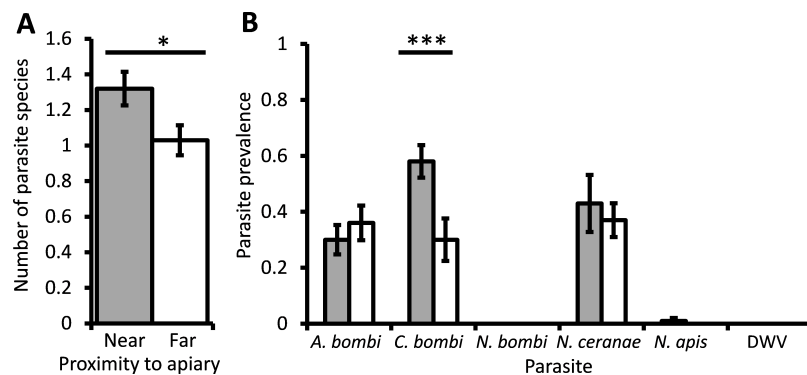


Figure 2 The effect of managed honeybees on parasite prevalence. The mean \pm s.e. parasite richness (number of species) per bumblebee (A), and the prevalence of six parasites per bumblebee colony (B), that were located either at a site near to (grey columns) or 1 km away from (white columns) a honey bee apiary. Asterisks indicate columns for which there was a significant difference between colonies located near to and far from the apiary (* $P < 0.05$; *** $P < 0.001$).

The effect of managed honey bees on parasite prevalence within bumblebee colonies

The mean parasite richness varied between bumblebee colonies but was significantly higher overall in colonies located in close proximity to honey bees ($\chi^2 = 5.66$, d.f. = 1, $P = 0.017$; Fig. 2A). The average prevalence of *C. bombi* in bumblebee colonies near honey bees was 58%; significantly higher than the 30% found in colonies far from honey bees ($\chi^2 = 17.9$, d.f. = 1, $P < 0.001$; Fig. 2B). The prevalence of *A. bombi* and *N. ceranae* in colonies located near honey bees averaged 30% and 43%, respectively, which did not differ from the prevalence of these parasites in colonies far from honey bees ($\chi^2 = 0.83$, d.f. = 1, $P = 0.36$; $\chi^2 = 0.27$, d.f. = 1, $P = 0.61$). *N. ceranae* prevalence did, however, differ between colonies within sampling sites ($\chi^2 = 25.07$, d.f. = 8, $P = 0.002$). *N. apis* was only found in bumblebee colonies located near to honey bee hives, but had a very low prevalence and thus did not differ significantly between the sites ($\chi^2 < 0.01$, d.f. = 1, $P = 0.993$). *Nosema bombi* and DWV were not detected in any of the 200 bumblebees sampled.

DISCUSSION

Although the study involved only a very limited number of sites and must thus be interpreted with caution, the results suggest that the prevalence of parasites in bumblebees may be affected by the presence of managed bees. The prevalence of *A. bombi* and *C. bombi* was respectively 12% and 15% higher in bumblebees near greenhouses at the three sites using commercially produced bumblebees compared to the two sites not using these bees, and the prevalence of *Apicystis bombi* was also much higher 0.5 km from the greenhouses compared with 5 km away from them. The samples were collected during a relatively short 16 day period and the differences between sites were not due to spatial or temporal autocorrelation. Bumblebees in colonies located close to the managed honey bee apiary had higher levels of the parasite *C. bombi* compared to bumblebees in colonies that were located 1 km away from the apiary. Although data from more sites are obviously needed to

draw firm conclusions, the results suggest that the presence of managed colonies of either bumblebees or honey bees may increase the prevalence of parasites in wild bumblebees.

A wide diversity of parasites were detected in the wild bumblebees collected near greenhouses, including the bumblebee parasites *A. bombi*, *C. bombi* and *N. bombi*, and the honey bee parasites *N. ceranae*, *N. apis* and DWV, all of which have also been identified in commercially produced bumblebees (Graystock et al., 2013b). *N. bombi*, *N. apis* and DWV were very rare (<1% prevalence) but the other parasites were more common. In general, the parasite richness within wild bumblebees increased with proximity to greenhouses utilising commercially produced bumblebees and bumblebees caught from around such greenhouses had a higher prevalence of *A. bombi* and *C. bombi* than those caught around greenhouses not using commercially reared bumblebees. Whether through parasite spillover, parasite spillback, or the stress of increased competition, commercially produced bumblebees appear to be increasing the prevalence of parasites in local bumblebees. These findings support previous, microscopy-based studies that found a higher prevalence of parasites near sites using commercially produced bumblebees (Colla et al., 2006; Murray et al., 2013; Otterstatter & Thomson, 2008). The effect of greenhouses using commercially produced bumblebees on the prevalence *A. bombi* appeared to be influenced by proximity to the focal glasshouse site. This perhaps suggests either a recent introduction from the greenhouses or that the dispersal of the parasite through the environment is relatively limited. There have been no studies of the horizontal transmission of *A. bombi*, although it has been commonly found at a low prevalence when bees are examined using less sensitive microscopy methods (Goulson, Whitehorn & Fowley, 2012; Shykoff & Schmid-Hempel, 1991). Worryingly this parasite has been implicated in bumblebee declines in South America (Arbetman et al., 2012). *Crithidia bombi* was also found to be more prevalent at sites using commercially produced bumblebees. Unlike *A. bombi*, there was no proximity effect found, but *C. bombi* is known to readily transmit between bumblebees and may therefore disperse rapidly through the environment (Durrer & Schmid-Hempel, 1994). The prevalence of none of the other parasites investigated differed between sites with or without commercially produced bumblebees. *Nosema ceranae* was abundant at some sites but completely absent at other sites. *Nosema ceranae*, is an emergent honey bee parasite that is implicated in the collapse of honey bee colonies in some, but not all, areas (Fries, 2010; Higes et al., 2008; Klee et al., 2007; Paxton, 2010; Paxton et al., 2008; Roberts & Hughes, 2014), and which has been shown to be widespread and virulent in bumblebees (Furst et al., 2014; Graystock et al., 2013a; Plischuk et al., 2009).

Although based on only two sites, the comparison of the site with honey bee hives and the site 1 km from the hives suggested that proximity to managed honey bee colonies may also have an effect on parasite prevalence in bumblebee colonies. Although the levels of *N. bombi*, *N. apis* and DWV were too low for any conclusions, and *A. bombi* and *N. ceranae* did not differ between the two sites, *C. bombi* was significantly more prevalent in bumblebee colonies that were near to the honey bee hives. This effect could not be due to spillover, because *C. bombi* is unable to infect honey bees (Ruiz-González & Brown, 2006). It could, however, be due to stress from competition leading to the bumblebees close to

the honey bee apiary being more susceptible to infection (*Brown, Loosli & Schmid-Hempel, 2000; Elbgami et al., 2014; Goulson & Sparrow, 2009; Lafferty & Gerber, 2002; Mallon, Brockmann & Schmid-Hempel, 2003*), or to the honey bees vectoring *C. bombi*. The potential role of stress and parasite spillback in driving elevated parasite prevalence in wild pollinators has been largely ignored and would warrant further investigation.

Our results suggest that managed colonies of bees may increase the prevalence of parasites in bumblebees. The results here are based on only very few sites and clearly further studies are needed using far more sites to establish their generality. It will be important for such studies to consider the potential for parasite spillback and stress-related effects, in addition to parasite spillover. It is clear that as long as there is mixing between managed and wild bees, there is the potential for wild populations to be at risk from the effects on host-parasite dynamics. These effects could prove to be a major conservation threat to bumblebees.

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Competing Interests

The authors declare there are no competing interests.

Author Contributions

- Peter Graystock conceived and designed the experiments, performed the experiments, analyzed the data, wrote the paper, prepared figures and/or tables.
- Dave Goulson conceived and designed the experiments, reviewed drafts of the paper.
- William O.H. Hughes conceived and designed the experiments, wrote the paper, reviewed drafts of the paper.

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REFERENCES

- Arbetman M, Meeus I, Morales C, Aizen M, Smagghe G. 2012. Alien parasite hitchhikes to Patagonia on invasive bumblebee. *Biological Invasions* 15:489–494 DOI 10.1007/s10530-012-0311-0.
- Arneberg P, Skorpung A, Grenfell B, Read AF. 1998. Host densities as determinants of abundance in parasite communities. *Proceedings of the Royal Society B: Biological Sciences* 265:1283–1289 DOI 10.1098/rspb.1998.0431.
- Biesmeijer JC, Roberts SPM, Reemer M, Ohlemuller R, Edwards M, Peeters T, Schaffers AP, Potts SG, Kleukers R, Thomas CD, Settele J, Kunin WE. 2006. Parallel declines in pollinators and insect-pollinated plants in Britain and the Netherlands. *Science* 313:351–354 DOI 10.1126/science.1127863.
- Brown MJF, Loosli R, Schmid-Hempel P. 2000. Condition-dependent expression of virulence in a trypanosome infecting bumblebees. *Oikos* 91:421–427 DOI 10.1034/j.1600-0706.2000.910302.x.
- Cameron SA, Lozier JD, Strange JP, Koch JB, Cordes N, Solter LF, Griswold TL. 2011. Patterns of widespread decline in North American bumble bees. *Proceedings of the National Academy of Sciences of the United States of America* 108:662–667 DOI 10.1073/pnas.1014743108.
- Chen YP, Higgins JA, Feldlaufer MF. 2005. Quantitative real-time reverse transcription-PCR analysis of deformed wing virus infection in the honeybee (*Apis mellifera* L.). *Applied and Environmental Microbiology* 71:436–441 DOI 10.1128/AEM.71.1.436-441.2005.
- Colla SR, Otterstatter MC, Gegear RJ, Thomson JD. 2006. Plight of the bumble bee: pathogen spillover from commercial to wild populations. *Biological Conservation* 129:461–467 DOI 10.1016/j.biocon.2005.11.013.
- Darvill B, Ellis JS, Lye GC, Goulson D. 2006. Population structure and inbreeding in a rare and declining bumblebee, *Bombus muscorum* (Hymenoptera: Apidae). *Molecular Ecology* 15:601–611 DOI 10.1111/j.1365-294X.2006.02797.x.
- Durrer S, Schmid-Hempel P. 1994. Shared use of flowers leads to horizontal pathogen transmission. *Proceedings of the Royal Society B: Biological Sciences* 258:299–302 DOI 10.1098/rspb.1994.0176.
- Elbgami T, Kunin WE, Hughes WOH, Biesmeijer JC. 2014. The effect of proximity to a honeybee apiary on bumblebee colony fitness, development, and performance. *Apidologie* 45(4):504–513 DOI 10.1007/s13592-013-0265-y.
- Ellis JS, Knight ME, Darvill B, Goulson D. 2006. Extremely low effective population sizes, genetic structuring and reduced genetic diversity in a threatened bumblebee species, *Bombus sylvarum* (Hymenoptera: Apidae). *Molecular Ecology* 15:4375–4386 DOI 10.1111/j.1365-294X.2006.03121.x.
- Evison SEF, Fazio G, Chappell P, Foley K, Jensen AB, Hughes WOH. 2013. Host-parasite genotypic interactions in the honey bee: the dynamics of diversity. *Ecology and Evolution* 3:2214–2222 DOI 10.1002/ece3.599.
- Evison SEF, Roberts KE, Laurenson L, Pietravalle S, Hui J, Biesmeijer JC, Smith JE, Budge G, Hughes WOH. 2012. Pervasiveness of parasites in pollinators. *PLoS ONE* 7:e30641 DOI 10.1371/journal.pone.0030641.
- Foley K, Fazio G, Jensen AB, Hughes WOH. 2012. Nutritional limitation and resistance to opportunistic *Aspergillus* parasites in honey bee larvae. *Journal of Invertebrate Pathology* 111:68–73 DOI 10.1016/j.jip.2012.06.006.
- Fries I. 2010. *Nosema ceranae* in European honey bees (*Apis mellifera*). *Journal of Invertebrate Pathology* 103:S73–S79 DOI 10.1016/j.jip.2009.06.017.

- Furst MA, McMahon DP, Osborne JL, Paxton RJ, Brown MJF. 2014. Disease associations between honeybees and bumblebees as a threat to wild pollinators. *Nature* 506:364–366 DOI 10.1038/nature12977.
- Gegear RJ, Otterstatter MC, Thomson JD. 2005. Does parasitic infection impair the ability of bumblebees to learn flower-handling techniques? *Animal Behaviour* 70:209–215 DOI 10.1016/j.anbehav.2004.09.025.
- Genersch E, Yue C, Fries I, de Miranda JR. 2006. Detection of deformed wing virus, a honey bee viral pathogen, in bumble bees (*Bombus terrestris* and *Bombus pascuorum*) with wing deformities. *Journal of Invertebrate Pathology* 91:61–63 DOI 10.1016/j.jip.2005.10.002.
- Gisder S, Genersch E. 2013. Molecular differentiation of *Nosema apis* and *Nosema ceranae* based on species-specific sequence differences in a protein coding gene. *Journal of Invertebrate Pathology* 113:1–6 DOI 10.1016/j.jip.2013.01.004.
- Goka K, Okabe K, Yoneda M, Niwa S. 2001. Bumblebee commercialization will cause worldwide migration of parasitic mites. *Molecular Ecology* 10:2095–2099 DOI 10.1046/j.0962-1083.2001.01323.x.
- Goulson D. 2003. *Bumblebees: behaviour and ecology*. Oxford: Oxford University Press.
- Goulson D, Hanley ME, Darvill B, Ellis JS, Knight ME. 2005. Causes of rarity in bumblebees. *Biological Conservation* 122:1–8 DOI 10.1016/j.biocon.2004.06.017.
- Goulson D, Lye GC, Darvill B. 2008. Decline and conservation of bumble bees. *Annual Review of Entomology* 53:191–208 DOI 10.1146/annurev.ento.53.103106.093454.
- Goulson D, Sparrow K. 2009. Evidence for competition between honeybees and bumblebees; effects on bumblebee worker size. *Journal of Insect Conservation* 13:177–181 DOI 10.1007/s10841-008-9140-y.
- Goulson D, Whitehorn P, Fowley M. 2012. Influence of urbanisation on the prevalence of protozoan parasites of bumblebees. *Ecological Entomology* 37:83–89 DOI 10.1111/j.1365-2311.2011.01334.x.
- Graystock P, Yates K, Darvill B, Goulson D, Hughes WOH. 2013a. Emerging dangers: deadly effects of an emergent parasite in a new pollinator host. *Journal of Invertebrate Pathology* 114:114–119 DOI 10.1016/j.jip.2013.06.005.
- Graystock P, Yates K, Evison SEF, Darvill B, Goulson D, Hughes WOH. 2013b. The Trojan hives: pollinator pathogens, imported and distributed in bumblebee colonies. *Journal of Applied Ecology* 50:1207–1215 DOI 10.1111/1365-2664.12134.
- Higes M, Martín-Hernández R, Botías C, Bailón EG, González-Porto AV, Barrios L, del Nozal MJ, Bernal JL, Jiménez JJ, Palencia PG, Meana A. 2008. How natural infection by *Nosema ceranae* causes honeybee colony collapse. *Environmental Microbiology* 10:2659–2669 DOI 10.1111/j.1462-2920.2008.01687.x.
- Kelly DW, Paterson RA, Townsend CR, Poulin R, Tompkins DM. 2009. Parasite spillback: a neglected concept in invasion ecology? *Ecology* 90:2047–2056 DOI 10.1890/08-1085.1.
- Klee J, Besana AM, Genersch E, Gisder S, Nanetti A, Tam DQ, Chinh TX, Puerta F, Ruz JM, Kryger P, Message D, Hatjina F, Korpela S, Fries I, Paxton RJ. 2007. Widespread dispersal of the microsporidian *Nosema ceranae*, an emergent pathogen of the western honey bee, *Apis mellifera*. *Journal of Invertebrate Pathology* 96:1–10 DOI 10.1016/j.jip.2007.02.014.
- Klee J, Tek Tay W, Paxton RJ. 2006. Specific and sensitive detection of *Nosema bombi* (Microsporidia: Nosematidae) in bumble bees (*Bombus* spp.; Hymenoptera: Apidae) by PCR of partial rRNA gene sequences. *Journal of Invertebrate Pathology* 91:98–104 DOI 10.1016/j.jip.2005.10.012.

- Kraus FB, Szentgyorgyi H, Rozej E, Rhode M, Moron D, Woyciechowski M, Moritz RFA. 2011. Greenhouse bumblebees (*Bombus terrestris*) spread their genes into the wild. *Conservation Genetics* 12:187–192 DOI 10.1007/s10592-010-0131-7.
- Lafferty KD, Gerber LR. 2002. Good medicine for conservation biology: the intersection of epidemiology and conservation theory. *Conservation Biology* 16:593–604 DOI 10.1046/j.1523-1739.2002.00446.x.
- Mallon EB, Brockmann A, Schmid-Hempel P. 2003. Immune response inhibits associative learning in insects. *Proceedings of the Royal Society B: Biological Sciences* 270:2471–2473 DOI 10.1098/rspb.2003.2456.
- Manson J, Otterstatter M, Thomson J. 2010. Consumption of a nectar alkaloid reduces pathogen load in bumble bees. *Oecologia* 162:81–89 DOI 10.1007/s00442-009-1431-9.
- Meeus I, Brown MJF, De Graaf DC, Smagghe GUY. 2011. Effects of invasive parasites on bumble bee declines. *Conservation Biology* 25:662–671 DOI 10.1111/j.1523-1739.2011.01707.x.
- Meeus I, Smagghe G, Siede R, Jans K, de Graaf DC. 2010. Multiplex RT-PCR with broad-range primers and an exogenous internal amplification control for the detection of honeybee viruses in bumblebees. *Journal of Invertebrate Pathology* 105:200–203 DOI 10.1016/j.jip.2010.06.012.
- Morandin LA, Laverty TM, Kevan PG, Khosla S, Shipp L. 2001. Bumble bee (Hymenoptera: Apidae) activity and loss in commercial tomato greenhouses. *Canadian Entomologist* 133:883–893 DOI 10.4039/Ent133883-6.
- Murray TE, Coffey MF, Kehoe E, Horgan FG. 2013. Pathogen prevalence in commercially reared bumble bees and evidence of spillover in conspecific populations. *Biological Conservation* 159:269–276 DOI 10.1016/j.biocon.2012.10.021.
- Oldroyd BP. 2007. What’s killing American honey bees? *PLoS Biology* 5:e168 DOI 10.1371/journal.pbio.0050168.
- Otterstatter M, Thomson J. 2007. Contact networks and transmission of an intestinal pathogen in bumble bee (*Bombus impatiens*) colonies. *Oecologia* 154:411–421 DOI 10.1007/s00442-007-0834-8.
- Otterstatter MC, Thomson JD. 2008. Does pathogen spillover from commercially reared bumble bees threaten wild pollinators? *PLoS ONE* 3:e2771 DOI 10.1371/journal.pone.0002771.
- Paxton RJ. 2010. Does infection by *Nosema ceranae* cause “Colony Collapse Disorder” in honey bees (*Apis mellifera*)? *Journal of Apicultural Research* 49:80–84 DOI 10.3896/IBRA.1.49.1.11.
- Paxton RJ, Klee J, Korpela S, Fries I. 2008. *Nosema ceranae* has infected *Apis mellifera* in Europe since at least 1998 and may be more virulent than *Nosema apis*. *Apidologie* 38:558–565 DOI 10.1051/apido:2007037.
- Plischuk S, Martín-Hernández R, Prieto P, Lucía M, Botías C, Meana A, Abrahamovich AH, Lange C, Higes M. 2009. South American native bumblebees (Hymenoptera: Apidae) infected by *Nosema ceranae* (Microsporidia), an emerging pathogen of honeybees (*Apis mellifera*). *Environmental Microbiology Reports* 1:131–135 DOI 10.1111/j.1758-2229.2009.00018.x.
- Potts SG, Biesmeijer JC, Kremen C, Neumann P, Schweiger O, Kunin WE. 2010. Global pollinator declines: trends, impacts and drivers. *Trends in Ecology & Evolution* 25:345–353 DOI 10.1016/j.tree.2010.01.007.
- Power AG, Mitchell CE. 2004. Pathogen spillover in disease epidemics. *American Naturalist* 164:S79–S89 DOI 10.1086/424610.

- Ricketts TH, Regetz J, Steffan-Dewenter I, Cunningham SA, Kremen C, Bogdanski A, Gemmill-Herren B, Greenleaf SS, Klein AM, Mayfield MM, Morandin LA, Ochieng A, Viana BF. 2008. Landscape effects on crop pollination services: are there general patterns? *Ecology Letters* 11:499–515 DOI 10.1111/j.1461-0248.2008.01157.x.
- Roberts KE, Hughes WOH. 2014. Immunosenescence and resistance to parasite infection in the honey bee, *Apis mellifera*. *Journal of Invertebrate Pathology* 121:1–6 DOI 10.1016/j.jip.2014.06.004.
- Rogerson PA. 2010. *Statistical methods for geography: a student's guide*. London: SAGE Publications Ltd.
- Ruiz-González MX, Brown MJF. 2006. Honey bee and bumblebee trypanosomatids: specificity and potential for transmission. *Ecological Entomology* 31:616–622 DOI 10.1111/j.1365-2311.2006.00823.x.
- Schmid-Hempel P. 2011. *Evolutionary parasitology: the integrated study of infections, immunology, ecology and genetics*. Oxford: Oxford University Press.
- Shykoff JA, Schmid-Hempel P. 1991. Incidence and effects of 4 parasites in natural populations of bumble bees in Switzerland. *Apidologie* 22:117–125 DOI 10.1051/apido:19910204.
- Singh R, Levitt AL, Rajotte EG, Holmes EC, Ostiguy N, van Engelsdorp D, Lipkin WA, de Pamphilis CW, Toth AL, Cox-Foster DL. 2010. RNA viruses in hymenopteran pollinators: evidence of inter-taxa virus transmission via pollen and potential impact on non-*Apis* hymenopteran species. *PLoS ONE* 5:e14357 DOI 10.1371/journal.pone.0014357.
- Szabo ND, Colla SR, Wagner DL, Gall LF, Kerr JT. 2012. Do pathogen spillover, pesticide use, or habitat loss explain recent North American bumblebee declines? *Conservation Letters* 5:232–239 DOI 10.1111/j.1755-263X.2012.00234.x.
- Vanbergen AJ, Baude M, Biesmeijer JC, Britton NF, Brown MJF, Brown M, Bryden J, Budge GE, Bull JC, Carvel C, Challinor AJ, Connolly CN, Evans DJ, Feil EJ, Garratt MP, Greco MK, Heard MS, Jansen VAA, Keeling MJ, Kunis WE, Marris GC, Memmott J, Murray JT, Nicolson SW, Osborne JL, Paxton RJ, Pirk CWW, Polce C, Potts SG, Priest NK, Raine NE, Roberts S, Ryabov EV, Shafr S, Shirley MDF, Simpson SJ, Stevenson PC, Stone GN, Termansen M, Wright GA, Insect Pollinators Initiative. 2013. Threats to an ecosystem service: pressures on pollinators. *Frontiers in Ecology and the Environment* 11:251–259 DOI 10.1890/120126.
- Velthuis HHW, van Doorn A. 2006. A century of advances in bumblebee domestication and the economic and environmental aspects of its commercialization for pollination. *Apidologie* 37:421–451 DOI 10.1051/apido:2006019.
- Whitehorn PR, Tinsley MC, Brown MJF, Darvill B, Goulson D. 2011. Genetic diversity, parasite prevalence and immunity in wild bumblebees. *Proceedings of the Royal Society B: Biological Sciences* 278:1195–1202 DOI 10.1098/rspb.2010.1550.
- Whittington R, Winston ML. 2003. Effects of *Nosema bombi* and its treatment fumagillin on bumble bee (*Bombus occidentalis*) colonies. *Journal of Invertebrate Pathology* 84:54–58 DOI 10.1016/S0022-2011(03)00123-X.
- Whittington R, Winston ML, Tucker C, Parachnowitsch AL. 2004. Plant-species identity of pollen collected by bumblebees placed in greenhouses for tomato pollination. *Canadian Journal of Plant Science* 84:599–602 DOI 10.4141/P02-192.